Nutritional indices, and proteolytic and digestive amylolytic activities of *Ephestia kuehniella* (Lep.: Pyralidae): response to flour of nine wheat cultivars

A. Abdi, B. Naseri* and S. A. A. Fathi

Department of Plant Protection, Faculty of Agricultural Sciences, University of Mohaghegh Ardabili, Ardabil, Iran. *Corresponding author, E-mail: bnaseri@uma.ac.ir

Abstract

The eggs and larvae of the Mediterranean flour moth, *Ephestia kuehniella* (Zell.) are widely used to rear parasitoids and predators for biological control programs. In this study, nutritional responses, proteolytic and digestive amylolytic activities of the fifth instar larvae as well as some biological parameters of *E. kuehniella* were studied on flour of nine wheat cultivars under laboratory conditions. The longest larval period of the fifth instar was on the cultivars Back cross Roshan and Sepahan and the shortest was on cultivar Tajan. The highest daily and total fecundity was observed on N-86-7. The fifth larval instar reared on Pishtaz showed the highest efficiency of conversion of ingested food. The highest general proteolytic activity was determined on the cultivars Kuhdasht and Khoshki line 9, respectively. According to the results obtained, Pishtaz and N-86-7 were the most suitable cultivars for laboratory rearing of *E. kuehniella* as the alternative host to use in the mass production of natural enemies.

Key words: Ephestia kuehniella, nutritional responses, proteolytic activity, amylolytic activity, wheat cultivars

چکیدہ

شاخصهای تغذیهای و فعالیت پروتئولیتیک و آمیلولیتیک گوارشی (Ephestia kuehniella (Lep.: Pyralidae: واکنش به آرد نه رقـم گندم

عا*ر*فه عبدی، بهرام ناصری و سید علی اصفر فتحی

تخمها و لاروهای شبپره مدیترانهای آرد، (.Zell) Ephestia kuehniella بهطور وسیعی در برنامههای کنترل بیولوژیک بهمنظور پروش پارازیتوییدها و شکارگرها مورد استفاده قرار میگیرند. در این تحقیق، واکنشهای تغذیهای، فعالیت پروتئولیتیک و آمیلولیتیک گوارشی لاروهای سن پنجم همراه با برخی پارامترهای زیستی E. kuehniella و رو آرد نه رقم گندم تحت شرایط آزمایشگاهی بررسی شدند. طولانی ترین دورهی لاروی سن پنجم روی ارقام Back cross Roshan و Roshan و کوتاه ترین آن روی رقم Tajan بود. بیش ترین باروری روزانه و کل روی رقم 8-86 مشاهده شد. لاروهای سن پنجم پرورشیافته روی رقم Pishta بیش ترین کارایی تبدیل غذای خورده شده را دارا بودند. بیش ترین فعالیت پروتئولیتیک کل روی رقم 7-86 و کم ترین آن روی رقم Pishta بیش ترین کارایی تبدیل غذای خورده شده را دارا بودند. بیش ترین فعالیت پروتئولیتیک کل روی رقم 7-86 و کم ترین آن روی رقم Pishta بیش ترین کارایی تبدیل غذای خورده شده را دارا بودند. بیش ترین فعالیت پروتئولیتیک کل روی رقم 7-86 و کم ترین آن روی رقم Pishta بیش ترین کارایی تبدیل غذای خورده شده را دارا بودند. بیش ترین فعالیت پروتئولیتیک کل روی رقم 7-86 و کم ترین آن روی رقم Pishta بیش ترین کارایی تبدیل غذای خورده شده را دارا بودند. بیش ترین فعالیت پروتئولیتیک کل روی رو مح و Pishta او Pishta بیش ترین کارایی تبدیل غذای دوره و که ترین فعالیت آمیلولیتیک گوارشی به تر تیب روی ارقام Kubasht و Pishta او Pishta او Pishta بود. Pishta رو Pishta و کم ترین فعالیت آمیلولیتیک گوارش و تر تولیتیک تر و ارقام Kubasht او Pishta او Pishta او Pishta بود. براساس نتایج به دست آمده، Pishta و 7-86 ارقام مناسبی برای پرورش آزمایشگاهی E. kuehniella و Pishta ای جریان جایگزین

واژگان کلیدی: Ephestia kuehniella واکنش های تغذیهای، فعالیت پروتئولیتیک، فعالیت آمیلولیتیک، ارقام گندم

Introduction

The Mediterranean flour moth, *Ephestia kuehniella* (Zell.), is one of the major pests of stored products, particularly flour (Sedlacek *et al.*, 1996; Hill, 2002; Rees, 2003), and is likely to be found in any mills or warehouses in which flour or other processed cereal products are stored (Sedlacek *et al.*, 1996). Although *E. kuehniella* is known as a destructive pest, the eggs and larvae of this species are widely utilized as an alternative host for the mass rearing of several parasitoids including *Habrobracon hebetor* (Say) (Hym.: Braconidae) (Shonouda & Nasr, 1998) and *Trichogramma ostriniae* (Peng & Chen) (Hym.: Trichogrammatidae) (Hoffmann *et al.*, 2001), and

predators including *Adalia bipunctata* (L.) (Col.: Coccinellidae) (Specty *et al.*, 2003), *Harmonia axyridis* Pallas (Col.: Coccinellidae) (De Clercq *et al.*, 2005), *Orius albidipennis* Reuter (Hem.: Anthocoridae) (Gonzalez-Zamora *et al.*, 2007), *Franklinothrips orizabensis* Johansen (Thys.: Aeolothripidae) (Hoddle *et al.*, 2001) and *Chrysoperla carnea* (Steph.) (Neu.: Chrysopidae) (Jokar & Golmohammadi, 2012; Jokar & Zarabi, 2012).

It is noticeable that species of *Trichogramma* Westwood are widely used in biological control against insect pests with over 15 million ha of agricultural crops being annually treated (van Lenteren, 2000; Wang *et al.*, 2014). The large numbers of *Trichogramma*, from parasitized eggs of stored product moths, are produced by commercial facilities (Bernardi *et al.*, 2000). Furthermore, it would be useful to store large numbers of adult host to achieve the acceptable number of host eggs when planning parasitoid releases. Also, the nutritional quality of host eggs and the subsequent survival of *Trichogramma* and other egg parasitoids depend on the diet of host (Hunter, 2003).

Several researchers have determined the effects of temperature and humidity (Siddiqui & Barlow, 1973), as well as the type of diet (Locatelli & Limonta, 1998) on the development of the Mediterranean flour moth. Locatelli et al. (2008) studied the effect of particle size of soft wheat flour on the development of E. kuehniella. Also, Pytelkova et al. (2009) investigated the molecular adaptation of amylolytic enzymes of E. kuehniella to alkaline environment. Recently, the midgut amylolytic activity of the Mediterranean flour moth was evaluated by Jafarlu et al. (2012), who reported the highest amylolytic activity in the 5th instar female larvae. Madboni & Pourabad (2012) considered the effect of different wheat cultivars on some of the developmental parameters of this pest, and showed that the fecundity of adults increased with increasing amounts of dietary protein.

Since wheat cultivars have different suitability as host to stored product insects (Amos et al., 1986; Sinha et al., 1988; McGaughey et al., 1990; Cortez-Rocha et al., 1993), and there is no published research article about nutritional responses, enzymatic activity and biological parameters of the Mediterranean flour moth on different wheat cultivars, therefore the present study was conducted to consider the proteolytic and digestive amylolytic activity, nutritional indices and life history aspects of E. kuehniella on the flour of different wheat cultivars. Due to the probable differences in nutritional values of wheat cultivars tested in this study and their effect on the enzymatic activity of E. kuehniella larvae, it was hypothesized that the larvae will accumulate biomass more efficiently when reared on some wheat cultivars than the others. It is further hypothesized that the adults who emerge from the larvae fed on the highnutritious cultivar will have high reproductive activity patterns. It is expected that the obtained results will be useful to introduce suitable wheat cultivar for the successful laboratory and mass rearing of *E. kuehniella* for its natural enemies.

Materials and methods

Wheat cultivars and insect rearing

Five wheat cultivars including Bam, Pishtaz, Sepahan, Khoshki line 9 and Back cross Roshan were obtained from Agricultural and Natural Resources Research Center of Isfahan, Iran. Four wheat cultivars including Tajan, Kuhdasht, N-86-7 and N-80-19 were obtained from Agricultural and Natural Resources Center, Moghan, Ardabil, Iran. Before starting the experiments, all wheat cultivars were milled and used with bran. The experiments were conducted during the morning to afternoon from July to December 2012.

The rearing stock of *E. kuehniella* was obtained from laboratory colony from Agricultural and Natural Resources Research Center of Isfahan, Iran. The larvae of *E. kuehniella* were reared on flour of nine wheat cultivars under a growth chamber at a temperature of 25 ± 1 °C, a relative humidity of $65 \pm 5\%$ and a 16:8 h light: dark photoperiod. The insects examined on different wheat cultivars had already been reared for two generations on the same cultivars.

Larval development and growth indices

Duration of last instar larvae, pupal period, their mortality, and the weights of larvae and pupae were recorded. Larval growth index (*LGI*) and standardized insect-growth index (*SII*) of *E. kuehniella* were calculated on different wheat cultivars using the following formulae (Itoyama *et al.*, 1999): $LGI = l_x/L$ and $SII = P_w/L$; where, l_x = survival rate of larvae, L = larval period and P_w = pupal weight.

Oviposition period, fecundity and longevity

After emergence of the adults, a pair of male and female moths (50 replicates for each cultivar) were transferred into egg-laying plastic cylinder (diameter 8 cm, depth 9 cm) covered by mesh net for aeration. The egg-laying containers were inversely placed on the paper sheets (as an oviposition surface), and the eggs were daily collected and counted. In the current research, the adults (males and females) were given neither food nor water (Xu *et al.*, 2008). Oviposition period, daily fecundity (eggs / reproduction day), total fecundity (eggs during the reproductive period) and the longevity of adults were recorded until the death of the last adult.

Nutritional indices

To start the experiment, 1 g of newly laid eggs was added in 750 g of wheat flour of each examined cultivar into plastic containers (diameter 20 cm, depth 8 cm) with a hole covered by a mesh net for aeration. Fifth instar larvae were collected from the containers and separated into five replicates (10 larvae in each) and transferred into plastic plate (diameter 8 cm, depth 1 cm), containing 1 g of flour of each examined cultivar. Nutritional indices were simply quantified using one-day-old fifth instar larvae as they were more measurable than the primary instars. The larvae were daily weighed, and the quantity of food ingested was daily calculated (by subtracting the diet remaining at the end of each experiment from the total weight of food given). To find the dry weights of the food and larvae, 100 g of wheat flour of the examined cultivars and 20 larvae that were reared on each cultivar were weighed, oven-dried (48 hours at 60 °C) and then reweighed to establish the percentage of their dry weight. The pupae were also weighed 24 hours after pupation.

Nutritional indices were calculated, based on dry weight, via formulae described by Waldbauer (1968): CI = E / A; ECI = P / E; $RCR = E / (A \times T)$; and RGR = $P / (A \times T)$; where, CI = consumption index, ECI = efficiency of conversion of ingested food, RCR = relative consumption rate, RGR = relative growth rate, A = mean dry weight of insect over unit time, E = dry weight of food consumed, P = dry weight gain of insect and T = duration of feeding period.

Chemicals

The general proteolytic substrate azocasein, Bradford reagent, the dinitrosalicylic acid (DNS), maltose and the amylolytic substrate starch were obtained from Sigma Chemical Co., St Louis, USA. Bovine serum albumin (BSA) and potassium iodine (KI) were purchased from Roche Co., and Merck Co., Germany, respectively. Iodine (I_2) was obtained from Maarssen Co., Netherlands.

Preparation of digestive enzymes

The rearing methods of larvae were similar to those mentioned in "insect rearing" section. After 12 h starvation, the whole body of the fifth instar larvae of *E. kuehniella*, which were reared for 24 h on flour of different wheat cultivars, were collected into a known volume of distilled water (after cold-immobilizing), and homogenized with a handheld glass grinder on ice. The homogenates were then centrifuged at $16000 \times g$ for 10 min at 4 °C and the resulting supernatants were collected in new micro tubes, stored at -20 °C in aliquots for further use.

Protein quantification

Protein concentrations in the fifth instar larvae of *E. kuehniella* were determined using bovine serum albumin (BSA) as standard according to the method of Bradford (1976).

Proteolytic activity assay

General proteolytic activity of one-day-old fifth instar larvae of *E. kuehniella* fed on different wheat cultivars was determined using azocasein substrate at the optimal pH. The universal buffer system (50 mM sodium phosphate borate) was used to assay the optimal pH of proteolytic activity (Elpidina *et al.*, 2001). To evaluate the azocaseinolytic activity, the reaction mixture containing 80 μ L of 1.5% azocasein solution in 50 mM universal buffer (pH 12) and 50 μ L of crude enzyme was incubated at 37 °C for 50 min. The reaction was terminated by the addition of 100 μ L 30% trichloroacetic acid (TCA), continued by cooling at 4 °C for 30 min and centrifugation at 16000 × g for 10 min. The supernatant (100 μ L) was added to 100 μ L of 2 M NaOH and the absorbance was read at 440 nm. Appropriate blanks, which TCA had been added prior to the substrate, were prepared for each treatment. One unit of protease activity was defined as an increase in optical density per milligram protein of the tissue min⁻¹ due to azocasein proteolysis (Elpidina *et al.*, 2001). All experiments were carried out in triplicates.

Amylolytic activity assay

Dinitrosalicylic acid (DNS) method (Bernfeld, 1955), with 1% soluble starch as substrate at the optimal pH was used to assay digestive amylolytic activity of the fifth instar larvae of *E. kuehniella* fed on different wheat cultivars. A quantity of 20 μ L of the enzyme extract was incubated with 500 μ L of universal buffer (pH 10) and 40 μ L of soluble starch for 30 min at 37 °C. The reaction was stopped by adding 100 μ L DNS and heating in boiling water for 10 min. The absorbance was read at 540 nm after cooling on ice. All experiments were carried out in triplicates.

Protein and starch determination of flour

Protein content of flour of wheat cultivars was quantified using BSA as standard according to Bradford (1976). A quantity of 200 mg of flour of each wheat cultivar was homogenized in 10 mL of distilled water, and then 100 µL of the homogenate was added to 3 mL of Bradford reagent. The samples were incubated in darkness at 37 °C for 60 min, and absorbance was read at 595 nm (Bouayad et al., 2008). Starch content of flour of wheat cultivars was determined by the method of Bernfeld (1955) using starch as standard. A quantity of 200 mg of flour of each wheat cultivar was homogenized in 35 mL of distilled water and heated to boiling point. One hundred microliters of each sample was added into 2.5 mL of iodine reagent (0.02% I_{2} and 0.2% KI) and absorbance was read at 580 nm (Bouayad et al., 2008).

Data analysis

Nutritional indices, enzymatic activity, and some biological parameters of *E. kuehniella* reared on flour of different wheat cultivars were analyzed using oneway ANOVA followed by comparison of the means with LSD test at $\alpha = 0.05$ using statistical software Minitab 16.0. All data were tested for normality before analysis. A dendrogram of nutritional indices, enzymatic activities and biological parameters of *E. kuehniella* on flour of different wheat cultivars was created after cluster analysis by Ward's method using SPSS 16.0 statistical software.

Results

Larval development and growth indices

The results showed that the larval period of the fifth instar was significantly different on nine wheat cultivars (F = 41.31; df = 8, 36; P < 0.01), the longest being on Back cross Roshan (12.00 ± 0.32 days) and Sepahan (12.00 \pm 0.45 days) and the shortest on Tajan $(5.60 \pm 0.40 \text{ days})$. The heaviest and the lightest larval weights (F = 5.63; df = 8, 36; P < 0.01) were observed on Sepahan (19.22 \pm 0.93 mg) and N-86-7 (14.05 \pm 0.89 mg), respectively. The longest pupal period (F =24.84; df = 8, 18; P < 0.01) was on Khoshki line 9 and Back cross Roshan (16.33 ± 0.33 days) and the shortest on N-80-19 (7.00 \pm 0.58 days). The heaviest and the lightest pupal weights (F = 61.07; df = 8, 36; P < 0.01) were detected in the larvae fed on N-86-7 (88.40 ± 4.04 mg) and Bam (12.82 \pm 0.16 mg), respectively. The results also showed that the highest larval growth index (LGI) was on cultivars Tajan, Kuhdasht and N-80-19 (16.66) and the lowest was on Sepahan (5.16). The standardized insect-growth index (SII) of E. kuehniella showed significant difference (F = 185; df = 8, 36; P <0.01) among the wheat cultivars, the highest being on N-86-7 (25.82 \pm 0.96 mg/day) and the lowest on Bam $(1.19 \pm 0.03 \text{ mg/day})$ (table 1).

Oviposition period, fecundity and longevity

Wheat cultivars tested showed significant effects on pre-oviposition period (F = 6.35; df = 8, 126; P < 0.01) of E. kuehniella, with the longest on Sepahan $(2.80 \pm 0.20 \text{ days})$ and the shortest on N-80-19 $(1.33 \pm$ 0.13 days). The longest oviposition period (F = 11.08; df = 8, 126; P < 0.01) on flour of different wheat cultivars was on Kuhdasht $(5.93 \pm 0.32 \text{ days})$, and the shortest on Pishtaz $(3.73 \pm 0.18 \text{ days})$. There was a significant difference in the effect of different cultivars of wheat as larval food on the number of eggs laid per reproduction day (F = 8.65; df = 8, 126; P < 0.01). The highest number of eggs laid per reproduction day was on N-86-7 (37.05 \pm 2.50 eggs), and the lowest one was on N-80-19 (14.30 \pm 0.93 eggs). There was also, a significant difference in the total number of eggs laid per individual of E. kuehniella on nine wheat cultivars (F = 11.18; df = 8, 126; P < 0.01), with the highest on N-86-7 (126.80 \pm 9.47 eggs) and the lowest on N-80-19 $(47.67 \pm 2.86 \text{ eggs})$ (table 2). Differences between the wheat cultivars showed no significant effect on the longevity of male E. kuehniella, but the longest female longevity (F = 12.86; df = 8, 126; P < 0.01) was on

Nutritional indices

The results of the nutritional responses of the fifth instar E. kuehniella larvae are shown in table 3. The highest food consumption (FC) (F = 15.40; df = 8, 36; P < 0.01) was observed in the larvae fed on N-80-19 $(72.91 \pm 6.30 \text{ mg/larva})$ and the lowest on Pishtaz (8.30 \pm 1.55 mg/larva). The highest and the lowest values of larval weight gain (F = 3.02; df = 8, 36; P < 0.05) were observed on N-80-19 (6.52 ± 1.08 mg) and Back cross Roshan (1.61 \pm 0.37 mg). Nutritional indices of the fifth instar larvae of E. kuehniella were significantly different on flour of wheat cultivars. The larvae reared on cultivars N-80-19 (9.83 \pm 2.14) and Pishtaz (0.48 \pm (0.08) showed the highest and the lowest values of CI (F =13.30; df = 8, 36; P < 0.01). The larvae reared on Pishtaz cultivar showed the highest value of ECI (F = 9.77; df = 8, 36; P < 0.01) (42.89 ± 9.19%), while the

Table 1. Duration of immature stages, larval and pupal weight and growth indices of *Ephestia kuehniella* on the flour of different wheat cultivars.

			Index (mean ± S	E)		
Wheat cultivar	Fifth instar larval period (day)	Larval weight (mg)	Pupal period (day)	Pupal weight (mg)	LGI	SII (mg/day)
Bam	10.80 ± 0.37 ab	16.54 ± 0.60 cd	7.33 ± 0.33 c	12.82 ± 0.16 e	8.80	1.19 ± 0.03 e
Pishtaz	10.60 ± 0.51 b	16.76 ± 0.46 cd	11.00 ± 0.58 b	26.47 ± 1.78 d	6.54	2.50 ± 0.19 de
Sepahan	12.00 ± 0.45 a	19.22 ± 0.93 a	14.33 ± 0.33 a	41.66 ± 1.68 c	5.16	3.57 ± 0.10 cd
Khoshki line 9	11.60 ± 0.25 ab	17.08 ± 0.31 bcd	16.33 ± 0.33 a	42.30 ± 1.38 c	5.33	3.52 ± 0.11 cd
Back cross Roshan	12.00 ± 0.32 a	18.96 ± 0.40 ab	16.33 ± 0.33 a	45.77 ± 2.35 c	5.83	3.80 ± 0.20 cd
Tajan	$5.60 \pm 0.51 \text{ d}$	15.33 ± 0.76 de	12.00 ± 1.15 b	26.72 ± 1.78 d	16.66	$4.91 \pm 0.55 \text{ c}$
Kuhdasht	$6.00 \pm 0.32 \text{ d}$	17.31 ± 0.74 abc	12.00 ± 0.58 b	61.36 ± 3.87 b	16.66	10.22 ± 0.65 b
N-86-7	7.40 ± 0.51 c	14.05 ± 0.89 e	12.00 ± 1.15 b	88.40 ± 4.04 a	10.00	25.82 ± 0.96 a
N-80-19	6.40 ± 0.51 cd	17.65 ± 0.78 abc	7.00 ± 0.58 c	60.50 ± 5.30 b	16.66	9.55 ± 1.04 b

The means followed by different letters in the same columns are significantly different (LSD, P < 0.01). LGI = larval growth index, SII = standardized insect-growth index.

Table 2. Oviposition period, fecundity and longevity of *Ephestia kuehniella* on the flour of different wheat cultivars.

	Index (mean ± SE)					
Wheat cultivar	Pre-oviposition period (day)	Oviposition period (day)	Daily fecundity	Total fecundity	Male longevity (day)	Female longevity (day)
Bam	1.73 ± 0.18 cd	5.73 ± 0.23 ab	18.47 ± 1.64 de	49.67 ± 4.25 ef	8.27 ± 0.15 a	5.73 ± 0.23 d
Pishtaz	1.67 ± 0.23 cd	$3.73 \pm 0.18 \text{ f}$	26.11 ± 2.91 bc	82.60 ± 6.13 cd	9.31 ± 0.22 a	7.13 ± 0.24 cd
Sepahan	2.80 ± 0.20 a	5.07 ± 0.28 bc	27.63 ± 3.31 bc	111.33 ± 11.00 ab	10.63 ± 0.49 a	8.33 ± 0.54 abc
Khoshki line 9	2.13 ± 0.27 bc	4.47 ± 0.29 cde	21.83 ± 2.85 cd	83.53 ± 10.20 cd	11.31 ± 0.25 a	9.60± 0.31 a
Back cross Roshan	$1.47 \pm 0.17 \text{ d}$	5.47 ± 0.24 ab	21.53 ± 2.10 cd	96.13 ± 12.50 bc	10.38 ± 0.30 a	8.60 ± 0.35 abc
Tajan	2.07 ± 0.12 c	3.80 ± 0.22 ef	22.75 ± 1.02 cd	70.73 ± 5.38 de	9.94 ± 0.27 a	7.80 ± 0.22 bc
Kuhdasht	1.80 ± 0.20 cd	5.93 ± 0.32 a	29.26 ± 1.72 b	102.53 ± 3.70 bc	11.63 ± 0.32 a	9.33 ± 0.29 ab
N-86-7	2.67 ± 0.25 ab	$4.13 \pm 0.22 \text{ def}$	37.05 ± 2.50 a	126.80 ± 9.47 a	10.13 ± 0.32 a	8.13 ± 0.35 abc
N-80-19	1.33 ± 0.13 d	4.67 ± 0.21 cd	14.30 ± 0.93 e	47.67 ± 2.86 f	9.13 ± 0.32 a	8.40 ± 0.27 abc

The means followed by different letters in the same columns are significantly different (LSD, P < 0.01).

lowest value was on Back cross Roshan (5.68 \pm 0.73%). The highest and the lowest values of *RCR* (F = 22.5; df = 8, 36; *P* < 0.01) were in the fifth instar larvae fed on N-80-19 (1.43 \pm 0.22 mg/mg/day) and Pishtaz (0.03 \pm 0.01 mg/mg/day) cultivars. Among different wheat cultivars, the *RGR* value (F = 2.57; df = 8; 36; *P* < 0.05) was the highest on Tajan (0.11 \pm 0.06 mg/mg/day) compared to other cultivars.

General proteolytic activity

General proteolytic activity data (P < 0.01) from *E. kuehniella* fifth instar larvae reared on flour of different wheat cultivars are indicated in table 4. The highest proteolytic activity of the fifth instars was in the larvae reared on N-86-7 (F = 79.80; df = 8, 18; P < 0.01) (1.98 ± 0.01 U mg⁻¹), while the lowest was on N-80-19 (1.10 ± 0.01 U mg⁻¹).

Amylolytic activity

Table 4 indicates digestive amylolytic activity (P < 0.01) from *E. kuehniella* fifth instar larvae reared on flour of different wheat cultivars. The larvae reared on Kuhdasht (F = 12.98; df = 8, 18; P < 0.01) showed the highest levels of amylolytic activity (2.22 ± 0.32 mU mg⁻¹), whereas the lowest activity was in the larvae reared on cultivar Khoshki line 9 (0.52 ± 0.05 mU mg⁻¹).

Protein and starch determination of flour

Statistical tests indicated significant differences in the protein and starch contents among flour of different wheat cultivars tested (P < 0.01) (table 5). The highest level of protein was in Sepahan (F = 35.71; df = 8, 18; P < 0.01) (1.571 ± 0.005 mg mL⁻¹), whereas the lowest level was in N-80-19 (1.474 ± 0.009 mg mL⁻¹). The highest and the lowest starch contents were found in flour of Kuhdasht (F = 261.34; df = 8, 18; P < 0.01) (0.368 ± 0.012 mg mL⁻¹) and Bam (0.032 ± 0.002 mg mL⁻¹), respectively (table 5).

Discussion

Growth, development, fecundity and physiological process of insects can be affected by the quality and the quantity of ingested food (Na & Ryoo, 2000; Musa & Ren, 2005). Current research demonstrated that different wheat cultivars had significant effect not only on the nutritional and enzymatic responses of *E. kuehniella*, but also on some of biological parameters of this insect.

The results showed that the larval period of the fifth instar was significantly different on nine wheat cultivars, which is in agreement with the findings of several researchers on other stored product moth pests. Locatelli & Limonta (1998) studied the development time of *E. kuehniella*, *Plodia interpunctella* (Hübner) and *Corcyra cephalonica* (Stainton) on different diets, and reported that the development time of these insects can be influenced by the different types of food consumed. In another study, Bouayad *et al.* (2008) demonstrated that the larval development time of *P. interpunctella* was significantly different on various food commodities.

Table 3. Nutritional resp	ses of the fifth instar larvae of <i>Ephestia kuehniella</i> on the flour of different wheat cul	ltivars.

	Index (mean \pm SE)					
Wheat cultivar	FC (mg/larva)	WG (mg)	CI	ECI (%)	RCR (mg/mg/day)	RGR (mg/mg/day)
Bam	13.38 ± 2.27 e	$3.73 \pm 0.72 \text{ bc}^*$	0.79 ± 0.11 c	34.37 ± 8.14 a	$0.07 \pm 0.01 \text{ d}$	$0.02 \pm 0.00 \text{ b}^*$
Pishtaz	8.30 ± 1.5 5e	4.77 ± 0.69 ab	$0.48 \pm 0.08 \text{ c}$	42.89 ± 9.19 a	$0.03 \pm 0.01 \text{ d}$	$0.02 \pm 0.01 \text{ b}$
Sepahan	32.58 ± 2.92 cd	2.75 ± 0.64 bc	1.79 ± 0.11 bc	$8.41 \pm 1.03 \text{ b}$	$0.13 \pm 0.01 \text{ d}$	$0.01 \pm 0.00 \text{ b}$
Khoshki line 9	32.85 ± 2.70 cd	3.26 ± 0.59 bc	2.01 ± 0.18 bc	10.75 ± 1.83 b	$0.14 \pm 0.01 \text{ d}$	$0.01 \pm 0.00 \text{ b}$
Back cross Roshan	29.58 ± 3.64 d	1.61 ± 0.37 c	1.56 ± 0.17 bc	5.68 ± 0.73 b	$0.11 \pm 0.01 \text{ d}$	$0.01 \pm 0.00 \text{ b}$
Tajan	45.89 ± 8.07 bc	4.34 ± 1.26 ab	3.02 ± 0.56 b	11.71 ± 1.78 b	0.59 ± 0.15 b	0.11 ± 0.06 a
Kuhdasht	32.48 ± 4.57 cd	2.79 ± 0.93 bc	1.89 ± 0.28 bc	10.15 ± 1.51 b	0.27 ± 0.04 cd	0.02 ± 0.01 b
N-86-7	47.44 ± 7.22 b	3.19 ± 0.70 bc	3.42 ± 0.54 b	7.45 ± 0.63 b	0.43 ± 0.07 bc	$0.03 \pm 0.01 \text{ b}$
N-80-19	72.91 ± 6.30 a	6.52 ± 1.08 a	9.83 ± 2.14 a	9.09 ± 1.56 b	1.43 ± 0.22 a	$0.03\pm0.01\ b$

The means followed by different letters in the same columns are significantly different (LSD, P < 0.01 and $P < 0.05^{\circ}$). FC = food consumed, WG = weight gain, CI = consumption index, ECI = efficiency of conversion of ingested food, RCR = relative consumption rate, RGR = relative growth rate.

The present study indicated that the heaviest and the lightest larval weights were on Sepahan and N-86-7, respectively (table 1). It would be concluded that the larvae fed on diet rich in protein content (Sepahan) showed the highest larval weight. It is generally recognized that the ultimate instars of lepidopteran larvae have greater needs for protein (Simpson et al., 1988), and the larvae fed on diets rich in protein demonstrate higher body weight (Sarate et al., 2012). Also, the pupal period and pupal weight of E. kuehniella were significantly affected by different wheat cultivars. The pupal period differed from 7.00 \pm 0.58 days on N-80-19 to 16.33 ± 0.33 days on cultivars Khoshki line 9 and Back cross Roshan, indicating that the differences in the nutrients or secondary compounds among the wheat cultivars can influence the pupal period. Leuck & Perkins (1972) noted that the pupal weight is an indicator of lepidopteran fitness. Since the heaviest pupal weight was in the larvae fed on N-86-7, and it has been previously reported that lepidopteran larvae fed on nutritive diets have a high pupal mass (Awmack & Leather, 2002), the highest standardized insect-growth index (25.82 ± 0.96 mg/day) was observed on this cultivar, indicating that N-86-7 was a nutritive wheat cultivar for feeding and growth of E. kuehniella. The results also showed that the highest value of the larval growth index was on Tajan, Kuhdasht and N-80-19, demonstrating that the larvae fed on these cultivars had higher survival rate than the larvae reared on other cultivars.

Because the types of diet consumed by the larval stages can influence the pre-oviposition and oviposition periods in the adult (Jallow *et al.*, 2001; Shayesteh *et al.*, 2010; Farahani *et al.*, 2011), the wheat cultivars tested in the current study showed significant effects on pre-oviposition and oviposition periods of *E. kuehniella*. Several authors have previously reported that the fecundity of moths can be affected by the types of different diets (Mbata, 1985; Kumral *et al.*, 2007; Fathipour & Naseri, 2011; Madboni & Pourabad, 2012; Arghand *et al.*, 2014). Accordingly, the fecundity of *E. kuehniella* was significantly influenced by the wheat

cultivars eaten by the larvae. Daily and total number of eggs per individual of the Mediterranean flour moth on nine wheat cultivars was highest on N-86-7, and lowest on N-80-19 (table 2). Since proteins are essential for egg production in the adult moth (Sorge et al., 2000), and cultivar N-80-19 was poor in protein content (table 5), the lowest fecundity was detected for the insects reared on this cultivar. Consequently, the highest fecundity detected on N-86-7 can be correlated with the highest pupal weight on this cultivar (Daryaei et al., 2007). Differences of wheat cultivars showed no significant effect on the longevity of male E. kuehniella, but the female longevity on different wheat cultivars differed from 5.73 ± 0.23 days on Bam to 9.60 ± 0.31 days on Khoshki line 9 (table 2). Higher fecundity of E. kuehniella reared on Khoshki line 9 than on Bam cultivars was probably attributed to longer female longevity on Khoshki line 9. Because the

Table 4. The mean $(\pm$ SE) proteolytic (U mg⁻¹) and digestive amylolytic (mU mg⁻¹) activities of whole body extracts from fifth instar larvae *Ephestia kuehniella* on the flour of different wheat cultivars.

Wheat cultivar	Proteolytic activity	Amylolytic activity		
Bam	1.61 ± 0.05 c	0.90 ± 0.13 de		
Pishtaz	$1.33 \pm 0.05 \text{ e}$	1.44 ± 0.17 bc		
Sepahan	$1.78 \pm 0.05 \text{ b}$	1.47 ± 0.07 bc		
Khoshki line 9	$1.30 \pm 0.01 \text{ e}$	$0.52 \pm 0.05 \text{ e}$		
Back cross Roshan	1.66 ± 0.04 c	$0.90 \pm 0.07 \text{ de}$		
Tajan	$1.46 \pm 0.00 \text{ d}$	1.05 ± 0.10 cd		
Kuhdasht	$1.81 \pm 0.00 \text{ b}$	2.22 ± 0.32 a		
N-86-7	1.98 ± 0.01 a	2.15 ± 0.12 a		
N-80-19	$1.10 \pm 0.01 \; f$	1.74 ± 0.23 ab		
The means followed by	different letters in t	the same columns a		

The means followed by different letters in the same columns are significantly different (LSD, P < 0.01).

Table 5. The mean $(\pm$ SE) protein and starch contents (mg mL⁻¹) of the flour of different wheat cultivars used for *Ephestia kuehniella* feeding.

Wheat cultivar	Protein content	Starch content
Bam	$1.548 \pm 0.003 \text{ b}$	0.032 ± 0.002 f
Pishtaz	$1.508 \pm 0.005 \text{ c}$	$0.166 \pm 0.003 \text{ d}$
Sepahan	1.571 ± 0.005 a	$0.236 \pm 0.010 \text{ c}$
Khoshki line 9	1.498±0.007 cd	$0.251 \pm 0.001 \text{ c}$
Back cross Roshan	1.493 ± 0.002 cde	$0.283 \pm 0.005 \text{ b}$
Tajan	1.482 ± 0.003 ef	$0.075 \pm 0.005 \text{ e}$
Kuhdasht	$1.492 \pm 0.004 \text{ de}$	0.368 ± 0.012 a
N-86-7	$1.500 \pm 0.005 \text{ cd}$	$0.147 \pm 0.008 \text{ d}$
N-80-19	$1.474 \pm 0.009 \text{ f}$	$0.149 \pm 0.004 \text{ d}$

The means followed by different letters in the same columns are significantly different (LSD, P < 0.01).

larvae fed on cultivar N-86-7 showed the highest pupal weight, standardized insect-growth index, and daily and total fecundity, this cultivar can be suggested as a suitable host diet for the growth and development of *E. kuehniella*.

Scanning the available literature revealed that the amounts and types of food eaten by an insect as well as the efficiency of conversion of ingested food are the factors determining the availability of nutrients for its maintenance and growth (Waldbauer, 1968; Slansky, 1990; Barton-Browne & Raubenheimer, 2003). The observed significant differences among nutritional indices, especially for ECI value, of E. kuehniella fed on the flour of different wheat cultivars indicating that the wheat cultivars had various nutritional values. It is noticeable that, among the nutritional indices, ECI is a feeding index that can be different due to variations in the food digestibility and the proportional amount of digestible food converted to insect biomass and metabolized to obtain the energy (Abdel-Rahman & Al-Mozini, 2007). Moreover, this index demonstrates an insect's ability to incorporate food into the growth (Nathan et al., 2005). Among the nine wheat cultivars, the highest CI value of E. kuehniella was on N-80-19 (table 3), indicating that the rate of intake relative to the mean larval weight during the feeding period was the highest on this cultivar. It is reported that the insect larvae effort to compensate for reduced ECI by increasing their CI (Price et al., 1980), a response that has been reported by many other researchers for the insect nutrition (Naseri et al., 2010; Mansouri et al., 2013; Rahimi Namin et al., 2014). In the current study, the highest ECI value of the Mediterranean flour moth was recorded for the larvae fed on cultivar Pishtaz, suggesting that they were more efficient at converting ingested food to body matter. The lowest ECI value on N-86-7 may be attributed to the deficiency of nutritional components and/or presence of some secondary chemicals in this cultivar. Nutritional requirements of an insect are correlated with body biomass and the period of immature stages. Furthermore, as the amount of food ingested is decreased, the insect becomes lighter (Lazarevic & Peric-Mataruga, 2003). Additionally, the significantly higher *ECI* achieved by the fifth instar larvae fed on Pishtaz suggests that the larvae fed on this cultivar had higher weight gain (tables 3). Insects consume less of a special diet simply because they are able to convert it more efficiently into the growth. Also, when the larvae consume less, the food will tend to pass through their alimentary canal slowly, and it can be efficiently converted into their body matter (Soo Hoo & Fraenkel, 1966). Accordingly, the lowest *RCR* value was observed in the last instar larvae of the Mediterranean flour moth fed on Pishtaz, while the highest value of *ECI* was on this cultivar (table 3).

It is well known that the soluble carbohydrates and proteins are efficiently utilized by insects and many species obtain most of their nourishment from these nutrients (Ishaaya et al., 1971). In the insects, the degree of food utilization has a direct impact on the enzymatic activity responsible for providing energy and nutrition to the growing larvae (Sivakumar et al., 2006), thus, for availability of the required nutrients, regulation of digestive enzymes is important (Kotkar et al., 2009). Proteases catalyze the release of amino acids from dietary proteins in an insect's alimentary canal to meet its nutritional necessities (Terra & Ferreira, 1994), and amylases play a main role in the starch digestion (Valencia-Jimenez et al., 2008). In the current research, general proteolytic and digestive amylolytic activities of E. kuehniella fifth instar larvae were studied. According to Razavi Tabatabaei et al. (2011), comparison of α -amylase activity in the whole body extracts from *Ectomyelois ceratoniae* (Zeller) (Lep.: Pyralidae) fifth instar larvae with the midgut extract indicated that all amylase activity from the larval body comes from the midgut. Therefore, it could be suggested that digestive amylolytic activity in E. kuehniella whole larval body extracts comes from the larval midgut. The present research demonstrated that the proteolytic and digestive amylolytic activities of the Mediterranean flour moth were significantly affected by the flour of different wheat cultivars (table 4). The highest proteolytic activity of the fifth instars was in the larvae reared on N-86-7; however, this cultivar had the moderate content of protein. The lowest proteolytic activity and protein content was detected on N-80-19 (tables 4 and 5). This is in agreement with the results of other researchers, who stated that the insects can release less of the digestive enzymes for nutrients present in excess, while maintaining or increasing the enzyme levels for nutrients in deficit (Kotkar et al., 2009; Lwalaba et al., 2010). Since protein intake occurs absolutely during the larval stages (Sorge et al., 2000), the larvae can not allow any dietary protein to pass undigested through the alimentary canal. So, it is helpful to keep a constant level of protease release in the event that even a small amount of protein is eaten (Lwalaba et al., 2010).

It is generally accepted that the insects can regulate the expression of digestive amylase regarding to the substrate levels of their food (Sarate et al., 2012). Because variations in the starch content in wheat cultivars can lead to differences in the amylolytic activity of E. kuehniella, thus, the highest amylolytic activity was detected in the larvae fed on Kuhdasht cultivar, which is attributed to the highest starch content of this cultivar. However, the larvae reared on Khoshki line 9 had the lowest levels of amylolytic activity, which was approximately four-fold lower than Kuhdasht. The mean digestive amylolytic activity of the fifth instar larvae of E. kuehniella on different wheat cultivars was approximately four- and five-fold higher than those reported for the last instar female and male larvae of this insect on wheat flour, respectively (Jafarlu et al., 2012). Possible reasons for this inconsistency can be because of the differences of wheat cultivars, variations in experimental conditions and genetic variations as a result of laboratory rearing of E. kuehniella. Comparison of the relationships between enzymatic activity and protein and starch contents of various cultivars tested demonstrated that there is likely an insect mechanism to exactly determine the diet contents, and regulate the levels of digestive enzymes (Kotkar et al., 2009). Higher protease and amylase activities of *E. kuehniella* larvae on some wheat cultivars can be because of the differences in either protein and starch contents of the food or presence of enzyme-inhibitors.

The soluble protein estimations of the flour of nine wheat cultivars indicated that Sepahan had the highest protein content. In the case of Pishtaz as a high-nutritious cultivar for *E. kuehniella*, the levels of protease and amylase activity are down-regulated to metabolize the requisite energy from ingested food. Because low dietary protein can cause a boosting in the consumption rate (Slansky, 1993); the highest food consumption of the fifth instar larvae of *E. kuehniella* was observed on cultivar N-80-19.

Comparison of the enzymatic activity, biological parameters and fecundity of the Mediterranean flour moth on the wheat cultivars tested revealed that the lowest daily and total fecundity as well as the highest values of CI and RCR were in the larvae reared on N-80-19, indicating that this cultivar is the least suitable wheat cultivar for rearing of E. kuehniella, however it can be utilized as a resistance cultivar to design new strategies for this pest management. Because the highest pupal weight, SII and fecundity (daily and total) were in E. kuehniella reared on N-86-7, and the larvae fed on Pishtaz showed the highest value of ECI, these two cultivars can be introduced as suitable wheat cultivars for feeding and rearing of E. kuehniella. For the future, it is recommendable to study the life table parameters and specific activity of digestive proteinases and carbohydrases of E. kuehniella in response to feeding on different wheat cultivars.

Acknowledgments

We gratefully appreciate M. R. Bagheri (University of Mohaghegh Ardabili, Ardabil, Iran) for his valuable assistance in this study. This work is financially supported by the University of Mohaghegh Ardabili (Ardabil, Iran), which is acknowledged. References

- Amos, T. G., Semple, R. L. & Williams, P. (1986) Multiplication of some stored grain insects on varieties of wheat. General and Applied Entomology 18, 48-52.
- Abdel-Rahman, H. R. & Al-Mozini, R. N. (2007) Antifeedant and toxic activity of some plant extracts against larvae of cotton leafworm *Spodoptera littoralis* (Lepidoptera: Noctuidae). *Pakistan Journal of Biological Sciences* 10, 4467-4472.
- Arghand, A., Naseri, B., Razmjou, J., Hassanpour, M. & Rahimi Namin, F. (2014) Comparative biological parameters of the cotton bollworm, *Helicoverpa armigera* (Lepidoptera: Noctuidae) on various corn hybrids. *Journal of Entomological Society of Iran* (in press).
- Awmack, C. S. & Leather, S. R. (2002) Host plant quality and fecundity in herbivorous insects. Annual Review of Entomology 47, 817-844.
- Barton-Browne, L. B. & Raubenheimer, D. (2003) Ontogenetic changes in the rate of ingestion and estimates of food consumption in fourth and fifth instar *Helicoverpa armigera* caterpillars. *Journal of Insect Physiology* 49, 63-71.
- Bernardi, E. B., Haddad, M. L. & Parra, J. R. P. (2000) Comparison of artificial diets for rearing *Corcyra cephalonica* (Stainton, 1865) (Lep.: Pyralidae) for *Trichogramma* mass production. *Revista Brasileira de Biologia* 60, 45-52.
- **Bernfeld**, **P.** (1955) Amylase, α and β . Methods in Enzymology 1, 149-154.
- Bouayad, N., Rharrabe, K., Ghailani, N. & Sayah, F. (2008) Effects of different food commodities on larval development and α-amylase activity of *Plodia interpunctella* (Hübner) (Lepidoptera: Pyralidae). *Journal of Stored Products Research* 44, 373-378.
- Bradford, M. A. (1976) Rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72, 248-254.
- Cortez-Rocha, M. O., Wong-Corral, F. J., Borboa-Flores, J., Sanchez-Marinez, R. I. & Cinco-Moroyoqui, F. J. (1993) A study on the susceptibility of wheat varieties to *Rhyzopertha dominica* F. *Southwestern Entomologists* 18, 287-291.
- Daryaei, G. M., Darvishi, S., Etebari, K. & Salehi, M. (2007) Host preference and nutrition efficiency of the gypsy moth, *Lymantria dispar* L. (Lepidoptera: Lymantriidae), on different poplar clones. *Turkish Journal of Agriculture* 32, 469-476.
- De Clercq, P., Bonte, M., Van Speybroeck, K., Bolckmans, K. & Deforce, K. (2005) Development and reproduction of *Adalia bipunctata* (Coleoptera: Coccinellidae) on eggs of *Ephestia kuehniella* (Lepidoptera: Phycitidae) and pollen. *Pest Management Science* 61, 1129-1132.
- Elpidina, E. N, Vinokurov, K. S., Gromenko, V. A., Rudenshaya, Y. A., Dunaevsky, Y. E. & Zhuzhikov, D. P. (2001) Compartmentalization of proteinases and amylases in *Nauphoeta cinerea* midgut. Archives of Insect Biochemistry and Physiology 48, 206-216.
- Farahani, S., Naseri, B. & Talebi, A. A. (2011) Comparative life table parameters of the beet armyworm, *Spodoptera exigua* (Hübner) (Lepidoptera: Noctuidae) on five host plants. *Journal of Entomological Research Society* 13(1), 91-101.
- Fathipour, Y. & Naseri, B. (2011) Soybean cultivars affecting performance of *Helicoverpa armigera* (Lepidoptera: Noctuidae). pp. 599-630 in Ng, T. B. (Ed.) *Soybean – biochemistry, chemistry and physiology*. 642 pp. In Tech, Rijeka, Croatia.

- Gonzalez-Zamora, J. E., Camunez, S. & Avilla, C. (2007) Effects of *Bacillus thuringiensis* Cry toxins on developmental and reproductive characteristics of the predator *Orius albidipennis* (Hemiptera: Anthocoridae) under laboratory conditions. *Environmental Entomology* 36, 1246-1253.
- Hill, D. S. (2002) Pests of stored foodstuffs and their control. 476 pp. Dordrecht, Kluwer Academic Publishers.
- Hoddle, M. S., Jones, J., Oishi, K., Morgan, D. & Robinson, L. (2001) Evaluation of diets for the development and reproduction of *Franklinothrips orizabensis* (Thysanoptera: Aeolothripidae). *Bulletin of Entomological Research* 91, 273-280.
- Hoffmann, M. P., Ode, P. R., Walker, D. L., Gardner, J., van Nouhuys, S. & Shelton, A. M. (2001) Performance of *Trichogramma ostriniae* (Hymenoptera: Trichogrammatidae) reared on factitious hosts including the target host, *Ostrinia nubilalis* (Lepidoptera: Crambidae). *Biological Control* 21, 1-10.
- Hunter, M. D. (2003) Effects of plant quality on the population ecology of parasitoids. *Agricultural and Forest* Entomology 5, 1-8.
- Ishaaya, I., Moore, I. & Joseph, D. (1971) Protease and amylase activity in larvae of the Egyptian cotton worm, Spodoptra littoralis. Journal of Insect Physiology 17, 945-953.
- Itoyama, K., Kawahira, Y., Murata, M. & Tojo, S. (1999) Fluctuations of some characteristics in the common cutworm, *Spodoptera litura* (Lepidoptera: Noctuidae) reared under different diets. *Applied Entomology and Zoology* 34, 315-321.
- Jafarlu, R., Farshbaf Pourabad, R. Valizadeh, M., Mohammadi, D. & Ziaei Madboni, M. A. (2012) Evaluation of midgut α-amylase activity in the Mediterranean flour moth, *Anagasta kuehniella* (Zeller, 1879) (Lep., Pyralidae). *Journal of Agricultural Science (University of Tabriz)* 22(3), 115-126. [In Persian].
- Jallow, M. F. A., Matsumura, M. & Suzuki, Y. (2001) Oviposition preference and reproductive performance of Japanese *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae). *Applied Entomology and Zoology* 36(4), 419-426.
- Jokar, M. & Golmohammadi, Gh. (2012) Investigation of population growth parameters of *Chrysoperla carnea* (Steph.) (Neuroptera: Chrysopidae) on egg of flour moth, *Anagasta kuehniella* (Zeller) (Lep.: Pyralidae) and compared with two artificial diets. *Proceedings of 20th Iranian Plant Protection Congress, Vol. I, Pests*, p. 743.
- Jokar, M. & Zarabi, M. (2012) Surveying effect kind of food on biological parameters on *Chrysoperla carnea* (Neuroptera: Chrysopidae) under laboratory conditions. *Egyptian Academic Journal of Biological Sciences* 5(1), 99-106.
- Kotkar, H. M., Sarate, P. J., Tamhane, V. A., Gupta, V. S. & Giri, A. P. (2009) Responses of midgut amylases of *Helicoverpa armigera* to feeding on various host plants. *Journal of Insect Physiology* 55, 663-670.
- Kumral, N. A., Kovanci, B. & Akbudak, B. (2007) Life tables of the olive leaf moth, *Palpita unionalis* (Hübner) (Lepidoptera: Pyralidae), on different host plants. *Journal of Biological and Environmental Sciences* 1(3), 105-110.
- Lazarevic, J. & Peric-Mataruga, V. (2003) Nutritive stress effects on growth and digestive physiology of *Lymantria dispar* larvae. *Yugoslav Medical Biochemistry* 22, 53-59.
- Leuck, D. B. & Perkins, W. D. (1972) A method of evaluating fall armyworm progeny reduction when evaluating control achieved by host plant resistance. *Journal of Economic Entomology* 65, 482-483.
- Locatelli, D. P. & Limonta, L. (1998) Development of *Ephestia kuehniella* (Zeller), *Plodia interpunctella* (Hübner) and *Corcyra cephalonica* (Stainton) (Lepidoptera: Pyralidae) on kernels and wholemeal flours of *Fagopyrum esculentum* (Moench) and *Triticum aestivum* (L.). *Journal of Stored Products Research* 3, 269-276.

- Locatelli, D. P., Limonta, L. & Stampini, M. (2008) Effect of particle size of soft wheat flour on the development of *Ephestia kuehniella* Zeller (Lepidoptera: Pyralidae). *Journal of Stored Products Research* 44, 269-272.
- Lwalaba, D., Hoffmann, K. H. & Woodring, J. (2010) Control of the release of digestive enzymes in the larvae of the fall armyworm, Spodoptera frugiperda. Archives of Insect Biochemistry and Physiology 73, 14-29.
- McGaughey, W. H., Speirs, R. D. & Martin, C. R. (1990) Susceptibility of classes of wheat grown in the United States to stored-grain insects. *Journal of Economic Entomology* 83, 1122-1127.
- Madboni, M. A. Z. & Pourabad, R. F. (2012) Effect of different wheat varieties on some of developmental parameters of Anagasta kuehniella (Lepidoptera: Pyralidae). Munis Entomology and Zoology 7(2), 1017-1022.
- Mansouri, S. M., Nouri Ganbalani, G., Fathi, S. A. A., Naseri, B. & Razmjou, J. (2013) Nutritional indices and midgut enzymatic activity of *Phthorimaea operculella* (Lepidoptera: Gelechiidae) larvae fed different potato germplasms. *Journal of Economic Entomology* 106, 1018-1024.
- Mbata, G. N. (1985) Some physical and biological factors affecting oviposition by *Plodia interpunctella* (Hübner) (Lepidoptera: Phycitidae). *Insect Science and its Application* 6, 597-604.
- Musa, P. D. & Ren, S. X. (2005) Development and reproduction of *Bemisia tabaci* (Homoptera: Aleyrodidae) on three bean species. *Insect Science* 12, 25-30.
- Na, J. H. & Ryoo, M. I. (2000) The influence of temperature on development of *Plodia interpunctella* (Lepidoptera: Pyralidae) on dried vegetable commodities. *Journal of Stored Products Research* 36, 125-129.
- Naseri, B., Fathipour, Y., Moharramipour, S. & Hosseininaveh, V. (2010) Nutritional indices of the cotton bollworm, *Helicoverpa armigera*, on 13 soybean varieties. *Journal of Insect Science* 10, Article 151, 1-14.
- Nathan, S. S., Chung, P. G. & Murugan, K. (2005) Effect of biopesticides applied separately or together on nutritional indices of the rice leaf older *Cnaphalocroccis medinalis*. *Phytoparasitica* 33, 187-195.
- Price, P. W., Bouton, C. E., Gross, P., Mc Pheron, B. A., Thompson, J. N. & Weis, A. E. (1980) Interactions among three trophic levels: influence of plants on interactions between insect herbivores and natural enemies. *Annual Review of Ecology and Systematics* 11, 41-65.
- Pytelkova, J., Hubert, J., Lepsik, H., Sobotnik, J., Sindelka, R., Krizkova, I., Horn, M. & Mares, M. (2009) Digestive α-amylases of the flour moth *Ephestia kuehniella* – adaptation to alkaline environment and plant inhibitors. *Federation of European Biochemical Societies Journal* 276, 3531-3546.
- Rahimi Namin, F., Naseri, B. & Razmjou, J. (2014) Nutritional performance and activity of some digestive enzymes of the cotton bollworm, *Helicoverpa armigera*, in response to seven tested bean cultivars. *Journal of Insect Science* 14, Article 93, 1-18.
- Razavi Tabatabaei, P., Hosseininaveh, V., Goldansaz, S. H. & Talebi, Kh. (2011) Biochemical characterization of digestive proteases and carbohydrases of the carob moth, *Ectomyelois ceratoniae* (Zeller) (Lepidoptera: Pyralidae). *Journal of Asia-Pacific Entomology* 14, 187-194.
- Rees, D. (2003) Insects of stored products. 181 pp. London, CSIRO Publishing.
- Sarate, P. J., Tamhane, V. A., Kotkar, H. M., Ratnakaran, N., Susan, N., Gupta, V. S. & Giri, A. P. (2012) Developmental and digestive flexibilities in the midgut of a polyphagous pest, the cotton bollworm, *Helicoverpa* armigera. Journal of Insect Science 12, Article 42, 1-16.
- Sedlacek, J. D., Weston, P. A. & Barney, R. J. (1996) Lepidoptera and Psocoptera. pp. 41-70 in Subramanyam, B. & Hagstrum, D. W. (Eds) *Integrated management of insects in stored products*. 432 pp. New York, Marcel Dekker.
- Shayesteh, N., Marouf, A. & Amir-Maafi, M. (2010) Some biological characteristics of the *Batrachedra amydraula* Meyrick (Lepidoptera: Batrachedridae) on main varieties of dry and semi-dry date palm in Iran. *Proceedings of* 10th International Working Conference on Stored Product Protection, pp. 151-155.

- Shonouda, M. L. & Nasr, F. N. (1998) Impact of larval-extract (kairomone) of *Ephestia kuehniella* Zell. (Lep., Pyralidae), on the behaviour of the parasitoid *Bracon hebetor* Say. (Hym., Braconidae). *Journal of Applied Entomology* 122, 33-35.
- Siddiqui, W. H. & Barlow, C. E. (1973) Population growth of Anagasta kuehniella (Lepidoptera: Pyralidae) at constant and alternating temperatures. Annals of the Entomological Society of America 66, 579-585.
- Simpson, S. J., Simmonds, M. S. J. & Blaney, W. M. (1988) A comparison of dietary selection behaviour in larval Locusta migratoria and Spodoptera littoralis. Physiological Entomology 13, 225-238.
- Sinha, R. N., Demianyk, C. J. & McKenzie, R. I. H. (1988) Vulnerability of common wheat cultivars to major stored product beetles. *Canadian Journal of Plant Science* 68, 337-343.
- Sivakumar, S., Mohan, M., Franco, O. L. & Thayumanavan, B. (2006) Inhibition of insect pest α-amylases by little and winger millet inhibitors. *Pesticide Biochemistry and Physiology* 85, 155-160.
- Slansky, F. (1990) Insect nutritional ecology as a basis for studying host plant resistance. *Florida Entomologist* 73, 359-378.
- Slansky, F. (1993) Xanthine toxicity to caterpillars synergized by allopurinol, a xanthine dehydrogenase oxidase inhibitor. *Journal of Chemical Ecology* 19, 2635-2650.
- Soo Hoo, C. F. & Fraenkel, G. (1966) The consumption, digestion, and utilization of food plants by a polyphagous insect, *Prodenia eridania* (Cramer). *Journal of Insect Physiology* 12, 711-730.
- Sorge, D., Nauen, R., Range, S. & Hoffmann, K. H. (2000) Regulation of vitellogenesis in the fall armyworm, Spodoptera frugiperda (Lepidoptera: Noctuidae). Journal of Insect Physiology 46, 969-976.
- Specty, O., Febvay, G., Grenier, S., Delobel, B., Piotte, C., Pageaux, J. F., Ferran, A. & Guillaud, J. (2003) Nutritional plasticity of the predatory ladybeetle *Harmonia axyridis* (Coleoptera: Coccinellidae): comparison between natural and substitution prey. *Archives of Insect Biochemistry and Physiology* 52, 81-91.
- Terra, W. R. & C. Ferreira (1994) Insect digestive enzymes: properties, compartmentalization and function. Comparative Biochemistry and Physiology 109, 1-62.
- Valencia-Jimenez, A., Arboleda, J. W., Lopez Avila, A. & Grossi-de-Sa, M. F. (2008) Digestive α-amylase from *Tecia solanivora* larvae (Lepidoptera: Gelechiidae): response to pH, temperature and plant amylase inhibitors. *Bulletin of Entomological Research* 98, 575-579.
- van Lenteren, J. C. (2000) Success in biological control of arthropods by augmentation of natural enemies. pp. 77-103 in Gurr, G. M. & Wratten, S. D. (Eds) *Biological control: measures of success*. 429 pp. Kluwer Academic Publishers, Dordrecht, The Netherlands.
- Waldbauer, G. P. (1968) The consumption and utilization of food by insects. *Advances in Insect Physiology* 5, 229-288.
- Wang, Z. Y., He, K. L., Zhang, F., Lu, X. & Babendreier, D. (2014). Mass rearing and release of *Trichogramma* for biological control of insect pests of corn in China. *Biological Control* 68, 136-144.
- Xu, J., Wang, Q. & He, X. Z. (2008) Emergence and reproductive rhythms of *Ephestia kuehniella* (Lepidoptera: Pyralidae). *New Zealand Plant Protection* 61, 277-282.

Received: 23 July 2013 Accepted: 5 February 2014