ACCESS

OPEN



Effect of cold storage on performance of *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae) reared on *Ephestia kuehniella* and *Ectomyelois ceratoniae*

Alieh Amini¹ ⁽¹⁾, Mojtaba Hosseini¹, Hussein Sadeghi¹ ⁽¹⁾ & Seyed Hossein Goldansaz² ⁽¹⁾

1- Department of Plant Protection, College of Agriculture, Ferdowsi University of Mashhad, Mashhad, Iran

- m.hosseini@um.ac.ir in https://orcid.org/0000-0001-6708-4657
- Sadeghin@um.ac.ir Inters://orcid.org/0000-0002-8329-2699

2- Department of Plant Protection, College of Agriculture and Natural Resources, University of Tehran, Karaj, Iran

⊠ goldansz@ut.ac.ir [™] *https://orcid.org/0000-0001-7574-7553*

Abstract. Trichogrammatid wasps are the most common natural enemies around the world used for biocontrol many important lepidoptran pests. Therefore, the availability of suitable storage methods for these parasitoids is valuable in biological control. In this study, the performance of Trichogramma brassicae reared on two different hosts and in two different cold storage periods was investigated. Wasp pupae formed on *Ephestia kuehniella* and *Ectomyelois* ceratoniae eggs, were kept in 10 °C for 30 days and then in 4 °C for 4 and 8 weeks. The effect of the treatment combinations on wasp's emergence rate, sex ratio and the number of female and male parasitoids in parental wasps (F_0) and their progeny (F_1) were estimated. In addition, adult wasp's shape (wingless and abnormality) and parasitism rate were investigated in parental wasps (F_0) and their progeny (F_1). The results of this study showed that emergence rate of T. *brassicae* in F_0 was significantly influenced by the interaction between host and cold storage period. The highest and lowest emergence rate were respectively found on *E. creatoniae* in control and E. kuehniella in 8 weeks cold storage period. Sex ratio and the number of female parasitoids in F_0 and F_1 were significantly influenced either by host or cold storage period, whereas interactive effect of these factors did not affect significantly sex ratio and the number of females in both generations. Also, wingless and abnormality of wasps in F_0 was not significantly influenced by any treatment combination. Parasitism rate of wasps reared on both hosts in F₁, significantly decreased by increasing in cold storage duration. Finally, the results indicated that the most of qualitative characteristics of wasps grown on E. ceratoniae, were clearly better than those of wasps grown on *E. kuehniella* both in F₀ and F₁.

Article History

Received: 01 October 2023 Accepted: 21 January 2024 Subject Editor: Yaghub Fathipour

Keywords: biological control, egg parasitoid, augmentative release, quality control, carob moth

Citation: Amini, A., Hosseini, M., Sadeghi, H. & Goldansaz, S. H. (2024) Effect of cold storage on performance of *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae) reared on *Ephestia kuehniella* and *Ectomyelois ceratoniae. J. Entomol. Soc. Iran* 44 (2), 129–139.

Introduction

Ectomyelois ceratoniae (Zeller) (Lepidoptera: Pyralidae), is the key pest of pomegranate orchards in Iran and many regions around the world and causes significant damage on nut and fruit products such as almonds, pistachios, pomegranates, stone and pomes fruits (Botha *et al.*, 2004; Azema & Mirabzadae, 2005). Several control methods, including sterile insect technique (Chakroun *et al.*, 2017), mating disruption (Mamay *et al.*, 2016), mass trapping (Mamay & Dağ, 2016), biological control (Memari *et al.*, 2016; Zougari *et al.*, 2021), mechanical control like fruit bagging and removal (Mamay, 2021) and chemical control (Warner *et al.*, 1990; Mnif *et al.*, 2013), have been examined to reduce carob moth damage in pomegranate orchards. However, the use of insecticides is not

© © © 2024 by Author(s), Published by the Entomological Society of Iran

This Work is Licensed under Creative Commons Attribution-Non Commercial 4.0 International Public License.

applicable because the larvae are protected from insecticides inside the fruit (Kishani-Farahani *et al.*, 2012). Mechanical and biological control methods particularly egg parasitoids of *Trichogramma* species are mostly used in Iran to control this pest (Ebrahimi, *et al.*, 1998; Moezipour, 2006).

Trichogramma brassicae Bezdenco has been proven to be the most widespread *Trichogramma* species in Iran and is commonly mass-reared for release in rice, corn, cotton, tomato, soybean fields and pomegranate orchards to control serious lepidopteran pests (Farrokhi *et al.*, 2010; Moezipour *et al.*, 2008). The success of biological control of mentioned pest depends not only on the time and the number of *Trichogramma* wasps released, but also on their quality (i.e. emergence, longevity, fecundity and searching capacity) (Bigler, 1994; Cerutti & Bigler, 1995; Dutton *et al.*, 1996; Van Lenteren & Tommasini, 2002).

Long-term storage of *Trichogramma* species is possible; especially cold-storage methods are common (Wang & Smith, 1996; Garcia *et al.*, 2002; Ayvaz *et al.*, 2008). Two main cold storage techniques have been used in mass rearing of *Trichogramma* spp. including with and without previous diapause induction (Greenberg *et al.*, 1996). Pre-storage (representing fall conditions: acclimation period) and cold-storage (representing winter conditions), cause long-term storage in *T. brassicae* with high emergence rate of adults (Rahimi-Kalde *et al.*, 2017; Cagnotti *et al.*, 2018). Acclimation of either 30 days at 10 °C or 24 days at 13 °C allowed *T. brassicae* immatures to develop with a lower mortality than those exposed directly at 5 °C (Lessard & Boivin, 2013).

Also, duration of exposure to low temperature has considerable effect on quality of biological agents (Denlinger & lee, 2010). For example, different storage periods of the wasp's pupae in 4 °C (1, 2, 3 and 4 weeks) had a significant effect on the adult emergence rate of *T. cacoeciae* Marchal, *T. evanescens* westwood and *T. brassicae* (Ozder, 2004). A significant reduction in adult emergence due to the lower cold-storage temperatures and longer duration, compared with the control group, was observed in these three species and other *Trichogramma* spp. (Jalali & Singh, 1992; Pitcher *et al.*, 2002; Ozder, 2004).

On the other hand, different hosts produce parasitoids wasps with different performance and efficiency (Muli et al., 2010; Farazmand, 2007). For example, when the *T. evanescens* reared on three various hosts (*Sitotroga cerealella* Olivier, *Ephestia kuehniella* Zeller and *Galleria mellonella* Linnaeus) for controlling cotton bollworm *Helicoverpa armigera* (Hübner), had different qualitative characteristics such as parasitism and emergence rates, fecundity and longevity (El-Wakeil, 2007).

Indeed, there are many studies which are separately focused on the effect of cold storage time or host's species on the performance and efficiency of *Trichogramma* wasps (Rahimi-Kaldeh *et al.*, 2017; Abbes *et al.*, 2020). However, there is little information for influence of interaction between cold storage period and host's species on the abilities of *Trichogramma* wasps. Thus, in the current study, we aimed to evaluate interactive effect of cold storage periods (control, 4 and 8 weeks) and two different hosts (*E. kuehniella* and *E. ceratoniae*) on *T. brassicae* qualitative characteristics both in parental and progeny generations.

Materials and methods

Insect culture

Ectomyelois ceratoniae larvae were collected from pomegranate orchards in Birjand (South- Khorasan Province, Iran). The larvae were transferred to container $(30 \times 20 \times 5 \text{ cm})$, containing pistachio powder (with low marketability and quality; about 50 larvae per unit). Containers kept in a big cage with net walls ($80 \times 80 \times 150$ cm) in room conditions set at $25 \pm 2 \text{ °C}$, $50 \pm 5\%$ (RH), and 16:8 (L:D) photoperiod (Ziaoddini *et al.*, 2011). After adults' emergence, the moths were allowed to fly in the cage for one day to get ready for mating. Then, moths were transferred to small cages for egg-laying. The pomegranate moths were reared for several generations in this condition.

The populations of *E. kuehniella* and *T. brassicae* were obtained from the colonies maintained at Insect Ecology and Pest Management Laboratory of Agriculture Faculty in Ferdowsi University of Mashhad. The parasitoids were reared on 24 hours old eggs of the *E. kuehniella*, in growth chamber ($25 \pm 1 \text{ °C}$, $65 \pm 5\%$ RH, 16:8 [L:D]).

Cold storage treatments

The eggs of both hosts (*E. kuehniella* and *E. ceratoniae*) were sterilized by exposing them to UV-C lamp (15 W Philips Holand) with a wavelength of 280 nm with a distance of 30 cm for 30 minutes (Nazeri *et al.*, 2015). Fifty numbers of 24-hours old eggs of *E. kuehniella* and *E. ceratoniae*, were separately sprinkled over a 20% honey solution layer on paperboard strips. Each strip was placed in glass vial (10×1 cm) with five pair of newly emerged parasitoid, *Trichogramma brassicae* and a droplet of honey solution for feeding parasitoids. Vials were plugged with cotton and held for one day at 25 ± 1 °C, 65-70% RH and 16:8 h (L:D) photoperiod. After 24 h, adult parasitoids were removed from glass vials. Strips containing the parasitized eggs were kept under rearing conditions as above mentioned until they reached the blackened stage (6th day of the pupal stage). The pupal stage was used because it showed better tolerance to cold storage compared to other embryonic stages in several *Trichogramma* spp. (Lopez & Morrison, 1980; Jalali & Singh, 1992; Abbes *et al.*, 2020). Then vials were kept in 10 °C in darkness for 30 days (as acclimation duration) (Rahimi-Kaldeh *et al.*, 2017)

Two different cold storage durations were examined: the glass vials were stored in the refrigerator at 4 ± 1 °C in darkness for 4 and 8 weeks, respectively (Ozder & Saglam, 2005; Ayvaz, *et al.*, 2008). Treatment combinations including two different host's eggs and two cold storage durations were replicated 20 times. At the end of each storage duration, vials were transferred into an incubator and maintained at standard rearing conditions until adult's emergence. Control groups were kept at standard rearing conditions without any cold storage duration in 20 repetitions. The effect of cold storage durations and different hosts on qualitative characteristics of parasitoids were estimated by determining the emergence rate (dividing the number of parasitized eggs presenting emergence holes by the total number of parasitized eggs as a percentage for each egg card), sex ratio, the number of female and male parasitoids, and wingless percentage for each vial.

Quality of progeny

The mated female wasps (24-h old) from each treatment combination (cold storage period and different hosts), were used to quality assessment of the wasp progeny. One female wasp was released in a glass vial with a card contains 20 eggs of *E. ceratoniae* or *E. kuehniella.* Vial was plugged with cotton and held for one day at 25 ± 1 °C, 65-70% RH and 16:8h (L:D) photoperiod. After 24 h, adult parasitoid was removed from the glass vial and strip containing the parasitized eggs was kept under standard rearing conditions until adult emergence. Then the emergence rate, sex ratio and parasitism percentage were estimated for each vial. This experiment had 20 repetitions for each treatment combination.

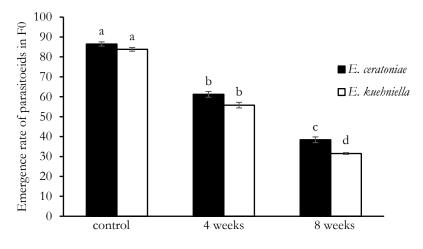
Statistical analyses

All data of wasps (F_0 and F_1) except wing deformity were tested for normality and the other assumptions of analysis of variance (ANOVA). The effect of cold storage periods and different hosts on qualitative characteristics of *T. brassicae* were analyzed by two-way full factorial ANOVA (cold storage period × different host). When ANOVA indicated a significant effect, means were separated using Tukey's honestly significant difference test. Wing deformity of wasps (F_0) was analyzed as binary responses using logistic regression (logit link function) with the cold storage period, different hosts and their interaction as predicted factors. All statistical analyses were performed using R project for statistical computing (R Core Team, 2020).

Results

Interactive effect of cold storage period and host on the emergence rate of adult wasps was significant in parental generation (F_0) ($F_{2, 114} = 13.8$, P < 0.01). The emergence rate of wasps stored on both hosts significantly decreased with an increase in cold storage period (Fig.1).

It was a significant reduction in sex ratio by increasing in cold storage period in F₀ (F_{2,114} = 55.7, P < 0.001), (Table 1). Also, the sex ratio of wasps was significantly higher on *E. ceratoniae* than that on *E. kuehniella* in F₀ (F_{1,114} = 15.85, P < 0.001), (Table 1). However, results showed that the interaction between host and cold storage period had no significant effect on sex ratio in F₀ (F_{2,114} = 0.44, P = 0.65). The number of female parasitoids was significantly affected either by cold storage period (F_{2,114} = 491.5, P < 0.001) or host (F_{1,114} = 36.14, P < 0.001), (Table 1).



cold storage period treatments

Fig 1. Emergence rate in the parental generation (F_0) of *Trichogramma brassicae* reared on *Ephestia kuehniella* and *Ectomyelois ceratoniae* in 0, 4 and 8 weeks cold storage periods. Different letters indicated a significant difference between treatment combinations (P < 0.01)

The number of male parasitoids was not significantly influenced neither by cold storage period ($F_{2,114} = 1.7$, P = 0.18) or host ($F_{1,114} = 2.14$, P = 0.11), (Table 1). The interaction between cold storage period and host did not affect the number of female and male parasitoids ($F_{2,114} = 1.7$, P = 0.19 and $F_{2,114} = 1.8$, P = 0.16, respectively).

Effect of cold storage period on wing deformity of parasitoid wasps in F_0 was not significant ($\chi^2 = 1.05$, df = 2, 116, P = 0.35). Similarly, there was no statistical differences in the number of adult parasitoid wasps with wing deformity among *E. ceratoniae* and *E. kuehniella* hosts ($\chi^2 = 1.41$, df = 1, 116, P = 0.25).

Either cold storage period or host affect significantly the emergence rate of *T. brassicae* wasps in progeny generation (F₁) (F_{2, 114} = 338.4, P < 0.001 and F_{1, 114} = 51.5, P < 0.001, respectively). However, the interaction between these factors did not affect this parameter (F_{2, 114} = 0.28, P = 0.75). Within the same host species, the emergence rate of wasps stored for 8 weeks, was significantly lower than those of stored for 4 weeks and control (Table 2). In comparison between two hosts' species, the emergence rate of adult wasps reared on eggs of *E. ceratoniae* was significantly higher than that reared on eggs of *E. kuehniella* (Table 2). Sex ratio of adult wasps in progeny generation (F₁) not only influenced by each factor individually (cold storage period: F_{2, 114} = 87.1, P < 0.001; host: F_{1, 114} = 4.7, P < 0.01) but also significantly affected by the interaction of both factors (F_{2, 114} = 3.6, P = 0.01). While the highest percent of sex ratio was observed from wasps kept on *E. ceratoniae* in control, the lowest percent of sex ratio was found from those stored on *E. kuehniella* in 8 weeks cold period (Table 2).

| I able 1. Effect of cold storage period and host on sex ratio, number of female and male of <i>I richogramma</i> |
|---|
| brassicae in parental generation F ₀ . Different letters in each column indicated a significant difference between |
| treatment combinations ($P < 0.01$), n = 20. |

| cold storage period | host | sex ratio (%) | number of females | number of males |
|---------------------|---------------|---------------|-------------------|-----------------|
| control | E. ceratoniae | 79.4±1.6a | 34.2±0.6a | 9.0±0.8a |
| | E. kuehniella | 70.0±3.4a | 29.2±1.4b | 12.7±1.6a |
| 4 weeks | E. ceratoniae | 65.6±1.8b | 19.9±0.5c | 10.6±0.7a |
| | E. kuehniella | 60±2.8b | 16.6±0.5d | 11.3±0.9a |
| 8 weeks | E. ceratoniae | 53.6±2.3c | 10.0±0.2e | 9.1±0.7a |
| | E. kuehniella | 44.5±1.7d | 7.6±0.2e | 9.6±0.3a |

| Parental cold storage period | host | emergence rate (%) | sex ratio (%) | number of female | number of male |
|---------------------------------|---------------|--------------------|---------------|------------------|----------------|
| control | E. ceratoniae | 67±1.3a | 76±2.0a | 9.1±0.3a | 2.8±0.2a |
| | E. kuehniella | 59±0.8b | 67±2.0b | 7.1±0.1b | 3.5±0.2a |
| 4 weeks | E. ceratoniae | 42±1.7c | 56±2.6b | 4.8±0.2c | 3.7±0.1a |
| | E. kuehniella | 34±1.1d | 54±2.2b | 3.8±0.1d | 3.1±0.2a |
| 8 weeks | E. ceratoniae | 28±0.8e | 40±0.02c | 2.2±0.1e | 3.3±0.1a |
| | E. kuehniella | 20±0.6f | 43±2.0c | 1.7±0.08e | 2.3±0.1b |

Table 2. Effect of cold storage period and host on emergence rate, sex ratio, number of female and male of *Trichogramma brassicae* in progeny generation F_1 . Different letters in each column indicated a significant difference between treatment combinations (P < 0.01), n = 20.

Interactive effect of cold storage period and host on the number of female and male parasitoids was also significant in progeny generation (F₁) (F_{2, 114} = 5.9, P < 0.01 and F_{2, 114} = 7.4, P < 0.01, respectively). The highest number of female parasitoids was found from wasps kept on *E. ceratoniae* in control, but the lowest ones was observed from those stored on *E. kuehniella* in 8 weeks cold period (Table 2).

The parasitism rate in progeny generation (F_1) not only was significantly influenced by cold storage period ($F_{2,114} = 148.5$, P < 0.001) and host ($F_{1,114} = 63.2$, P < 0.001) individually, but also it was significantly affected by the interaction between these two factors ($F_{2,114} = 4.1$, P = 0.02). The female wasps grown on the host with longer cold storage period had a significantly lower parasitism rate compare to those grown on the hosts with shorter cold storage period and control (Fig. 2). Also, the parasitism rate of female wasps grown on *E. ceratoniae* was significantly higher than those on *E. kuehniella*.

Discussion

It would be useful to store the parasitoid for some biocontrol program when synchronizing the parasitoids with pest population. However, the potential value for use of cold storage in commercial production needs to be evaluated in terms of economies. Finding methods to increase duration of storage for biological control agents is an important issue and helps producers to provide a large number of beneficial insects at the appropriate time in field. It also causes the easier acceptance of natural hosts by a reduction in the number of generations which is necessary for their mass rearing (Nazeri *et al.*, 2015; Rahimi-Kaldeh *et al.*, 2017).

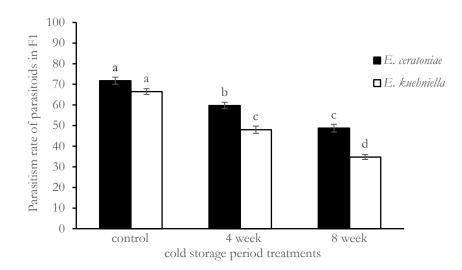


Fig 2. Parasitism rate in the progeny generation (F_1) of *Trichogramma brassicae* reared on *Ephestia kuehniella* and *Ectomyelois ceratoniae* in 0, 4 and 8 weeks cold storage periods. Different letters indicated a significant difference between treatment combinations (P < 0.01).

Several studies showed that temperature, photoperiod, cold storage duration, species or even strains of wasps and hosts, influence on diapause induction, post-diapause duration and quality of stored *Trichogramma* spp. (Laing & Corrigan, 1995; Rahimi-Kaldeh *et al.*, 2017). In our study, a reduction in the emergence rate of parasitoids due to the cold storage of *T. brassicae* was found and the percentage of parasitoid emergence rate in control was greater than any cold-stored ones. The decrease in the emergence rate of parasitoids primarily is due to the mortality caused by the cold storage of *T. brassicae* during the pupal stage and, due to negative physiological and physical changes that have been occurred to pupa of *T. brassicae*. Moreover, freezing and subsequent thawing could also have provoked changes in the hardness of the host eggs chorion and making it more difficult for the wasps to cut it with their mouth parts for emergency (Kivan Kilic, 2005). Unlike to the results of current study, 1 to 16 weeks cold storage of *T. bactrae* in 4 °C, had no significant effect on the emergence rate of female wasps (Mohamed & El-Heneidy, 2020), probably because of difference parasitoid species.

Here, cold storage specifically in a longer period (8 weeks) affected negatively the quality of emerged adults of *T. brassicae*, so that, the sex ratio and parasitism rate of *T. brassicae* lessened as cold storage period increased. Several studies have addressed the negative effect of cold storage period on the quality of emerged parasitoid adults (Tezze & Betto, 2004; Lessard & Bovin, 2013; Rahimi-Kaldeh *et al.*, 2017; Vaez *et al.*, 2018; Abbes *et al.*, 2020). These may be attributed to the increase in energy necessity and gradual weakness of the individual during metamorphosis in the storage period, and therefore accumulation of toxic metabolites and oxidative stress caused by a buildup of reactive oxygen (Amice *et al.*, 2008). The results also showed that the number of emerging female parasitoids significantly decreased with an increase in the cold storage period, while the number of emerging males was not affected by the period of cold storage in both the F₀ and F₁ generations. Some studies have reported a shift towards the production of male-biased parasitoids after prolonged cold exposure, indicating differential mortality of females (Farid *et al.*, 2001; Chen *et al.* 2008). The quality of the host influences the oviposition patterns of parasitoids, with female eggs typically being laid on higher-quality hosts (Van den Assem, 1971; Otto & Mackauer, 1998). Since the extended period of cold storage decreases the quality of the host (Colinet *et al.*, 2006), increasing the duration of host storage probably reduces the proportion of female offspring of parasitoids.

In some *Trichogramma* species and other parasitoids, morphological alterations occur during cold storage period; such as adult size reduction, deformation in wings, legs, antennae and sensillae structure (Pintureau and Daumal, 1995; Rundle *et al.*, 2004; Bourdais *et al.*, 2006), although our results didn't show a significant difference in adult deformation after cold storage in different host and cold storage period.

This study showed that *T. brassicae* pupae reared on *E. ceratoniae* eggs, were relatively amenable to cold storage and can be used in mass production process. The qualitative characteristics of F_0 and F_1 parasitoids reared on *E. ceratoniae*, were significantly better than those reared on *E. kuehniella*. This phenomenon may be attributed to that *E. ceratoniae* is natural host for *T. brassicae*, whereas *E. kuehniella* is a fictitious host for it.

In general, our results indicated that T. brassicae could be exposed to an additional 4 weeks at 4 °C after acclimation and diapause induction at 10 °C for 30 days without much decrease in fitness of parasitoids. Therefore, acclimation can protect T. brassicae efficiency and quality, and should be considered in storage protocol of this parasitoid. Similar results were reported by Rahimi-Kaldeh et al. (2017). The possible reason of acclimation during cold storage may be that the amount of cryoprotectants (e.g. glycerol and alanine), which reduce the ice formation at any temperature by increasing the total concentration of all the solutes, increases in diapausing insects during acclimation period (like autumn) and so, causes more adaptation for wasp pupa to tolerate cold storage period (Rivers et al., 2000). From a practical point of view, our results revealed that the treatment with 30 days of acclimation and 4 weeks of cold store is useful to establish a cold storage protocol for T. brassicae. However, the decrease of *T. brassicae* performance after cold storage period should be considered in mass production, and the release percentage should be increased by the equivalent of lack % emergence. Moreover, using the other methods such as 'recurrent warming method' and 'rapid acclimation method' [a short pre-exposure (minutes or hours) to sub-lethal low temperature] which are improve the efficiency of cold stored parasitoids, should be considered (Denlinger & Lee, 2010; Gardner et al., 2012). On the other hand, the influence of cold storage conditions on the field success of *T. brassicae* should be assessed because the efficacy of *Trichogramma* species is different in field from laboratory (Hached et al., 2021).

Acknowledgments

The authors are grateful to the Ferdowsi University of Mashhad for providing the facilities.

Funding

This research was supported by a grant from the Ferdowsi University of Mashhad (Project Number, P3/53874)

REFERENCES

- Abbes, K., Zouba, A., Harbi, A. & Chermiti, B. (2020) Effect of cold storage on the performance of *Trichogramma bourarachae* (Pintureau and Babault)(Hymenoptera: Trichogrammatidae). *Egyptian Journal of Biological Pest Control* 30, 1-6. https://doi.org/10.1186/s41938-020-00232-1
- Amice, G., Vernon, P., Outreman, Y., Van Alphen, J. & Van Baaren, J. (2008) Variability in responses to thermal stress in parasitoids. *Ecological Entomology* 33(6), 701-708. https://doi.org/10.1111/j.1365-2311.2008.01019.x
- Ayvaz, A., Karasu, E., Karabörklü, S. & Tunçbilek, A. Ş. (2008) Effects of cold storage, rearing temperature, parasitoid age and irradiation on the performance of *Trichogramma evanescens* Westwood (Hymenoptera: Trichogrammatidae). *Journal of Stored Products Research* 44(3), 232-240. https://doi.org/10.1016/j.jspr.2008.02.001
- Azema, M. & Mirabzadae, A. (2005) Issues on different aspects of applying natural enemies for biological control of insect pests. Sepehr Publication, Iran.
- **Bigler, F.** (1994) Quality control in *Trichogramma* production. In: Wajnberg E., & Hassan S. A. (eds.) Biological control with egg parasitoids. CAB International, Wallingford, CT. pp 87-115.
- Botha, J., Hardie, D. & Hoffmann, H. (2004) Carob moth. WA Department of Agriculture Garden Note, 21.
- Bourdais, D., Vernon, P., Krespi, L., Le Lannic, J. & Van Baaren, J. (2006). Antennal structure of male and female Aphidius rhopalosiphi DeStefani–Peres (Hymenoptera: Braconidae): description and morphological alterations after cold storage or heat exposure. Microscopy Research and Technique 69, 1005-1013. https://doi.org/10.1002/jemt.20378
- Cagnotti, C. L., Lois, M., López, S. N., Botto, E. N. & Viscarret, M. M. (2018) Cold storage of *Trichogramma nerudai* using an acclimation period. *BioControl* 63(4), 565-573. https://doi.org/10.1007/s10526-018-9885-5
- Cerutti, F. & Bigler, F. (1995) Quality assessment of *Trichogramma brassicae* in the laboratory. *Entomologia Experimentalis et Applicata* 75(1), 19-26. https://doi.org/10.1111/j.1570-7458.1995.tb01905.x
- Chakroun, S., Rempoulakis, P., Lebdi-Grissa, K. & Vreysen, M. J. (2017) Gamma irradiation of the carob or date moth *Ectomyelois ceratoniae*: dose-response effects on egg hatch, fecundity, and survival. *Entomologia Experimentalis et Applicata* 164(3), 257-268. https://doi.org/10.1111/eea.12617
- Chen, W. L., Leopold, R. A. &Harris, M. O. (2008) Cold storage effects on maternal and progeny quality of Gonatocerus ashmeadi Girault (Hymenoptera: Mymaridae). Biological Control 46, 122-132. https://doi.org/10.1016/j.biocontrol.2008.04.007
- Colinet, H., Hance, T. & Vernon, P. (2006) Water relations, fat reserves, survival, and longevity of a cold-exposed parasitic wasp *Aphidius colemani* (Hymenoptera: Aphidiinae). *Environmental Entomology* 35, 228-236. https://doi.org/10.1603/0046-225X-35.2.228
- Denlinger, D. L. & Lee Jr, R. E. (2010) Low temperature biology of insects. Cambridge University Press. http://dx.doi.org/10.1017/CBO9780511675997.
- Dhouibi, M. H., Hawari, W., Touil, S., Zouba, A. & Ben Hamida, F. (2017) Biocontrol of the carob moth *Ectomyelois ceratoniae* (Lepidoptera, Pyralidae) in two oases in the south of Tunisia using mating disruption with SPLAT EC O. *International Journal of Agriculture Innovations and Research* 5, 825-830.
- Dutton, A., Cerutti, F. & Bigler, F. (1996) Quality and environmental factors affecting *Trichogramma brassicae* efficiency under field conditions. *Entomologia experimentalis et applicate*, 81(1), 71-79. https://doi.org/10.1111/j.1570-7458.1996.tb02016.x
- Ebrahimi, E., Pintureau, B. & Shojai, M. (1998) Morphological and enzymatic study of the genus *Trichogramma* in Iran (Hym. Trichogrammatidae). *Applied Entomology and Phytopathology* 66(1-2), 39-40.
- El-Wakeil, N. E. (2007) Evaluation of efficiency of *Trichogramma evanescens* reared on different factitious hosts to control *Helicoverpa armigera. Journal of Pest Science* 80, 29-34. https://doi.org/10.1007/s10340-006-0150-9

- Farazmand, A., Iranipour, S. H., Saber, M. & Mashhadi Jafarloo, M. (2007) Comparison of some biological parameters of *Trichogramma brassicae* Bez. on *Anagasta kuehniella* Zell. and *Plodia interpunctella* (Hbn.) in laboratory condition. *Agriculture Science* 17, 175-185.
- Farid, A., Ullah, T., Khan, A., Khattak, S. U. & Pak, A. (2001) Effect of storage at low temperature on adult eclosion and longevity of adults of *Trichogramma chilonis. Pakistan Journal of Zoology* 33, 205-207.
- Farrokhi, S., Ashouri, A., Shirazi, J., Allahyari, H. & Huigens, M. E. (2010) A comparative study on the functional response of *Wolbachia*-infected and uninfected forms of the parasitoid wasp *Trichogramma brassicae*. *Journal of Insect Science* 10(1), 167. http://doi.org/10.1673/031.010.14127
- Garcia, P. V., Wajnberg, E., Pizzol, J. & Oliveira, M. L. M. (2002) Diapause in the egg parasitoid *Trichogramma* cordubensis: role of temperature. *Journal of Insect Physiology* 48(3), 349-355. https://doi.org/10.1016/S0022-1910(02)00052-5
- Gardner, J., Hoffmann, M. P., Pitcher, S. A. & Nyrop, J. P. (2012) Recurrent warming to improve cold storage of *Trichogrammatids* (Hymenoptera: Trichogrammatidae). *Biocontrol Science and Technology* 22(3), 261-270. https://doi.org/10.1080/09583157.2011.651099
- Greenberg, S. M., Nordlund, D. A. & King, E. G. (1996) Mass production of *Trichogramma* spp.: experiences in the former Soviet Union, China, the United States and Western Europe. *Biocontrol News and Information* 17, 51–60.
- Hached, W., Sahraoui, H., Blel, A. & Lebdi Grissa, K. (2021) Biological control of *Ectomyelois ceratoniae* Zeller (Lepidoptera: Pyralidae) using the egg parasitoid *Trichogramma cacoeciae* Marchal (Hymenoptera: Trichogrammatidae) in a Tunisian citrus orchard. *Journal of Entomology and Zoology Studies* 9(3), 98-104. https://www.entomoljournal.com/archives/?year=2021&vol=9&issue=3&ArticleId=8699
- Hérard, F., Keller, M. A., Lewis, W. J. & Tumlinson, J. H. (1988) Beneficial arthropod behavior mediated by airborne semiochemicals: III. Influence of age and experience on flight chamber responses of *Microplitis demolitor* Wilkinson. *Journal of Chemical Ecology* 14, 1583-1596. https://doi.org/10.1007/BF01012524
- Ivanov, M. F. & Reznik, S. Y. (2008) Photoperiodic regulation of the diapause of the progeny in *Trichogramma embryophagum* Htg. (Hymenoptera, Trichogrammatidae): Dynamics of sensitivity to photoperiod at the immature stages of maternal females. *Entomological Review* 88(3), 261-268. https://doi.org/10.1134/S0013873808030019
- Jalali, S. K. & Singh, S. P. (1992) Differential response of four *Trichogramma* species to low temperatures for short term storage. *Entomophaga* 37, 159-165. https://doi.org/10.1007/BF02372984
- Kishani-Farahani, H., Goldansaz, S. H. & Sabahi, Q. (2012) A survey on the overwintering larval parasitoids of *Ectomyelois ceratoniae* in three regions in Iran. *Crop Protection* 36, 52-57. https://doi.org/10.1016/j.cropro.2012.01.018
- Kivan, M. & Kilic, N. (2005) Effects of storage at low-temperature of various heteropteran host eggs on the egg parasitoid, *Trissolcus semistriatus. Biocontrol* 50, 589-600. https://doi.org/10.1007/s10526-004-3122-0
- Koštál, V. (2006) Eco-physiological phases of insect diapause. *Journal of Insect Physiology* 52(2), 113-127. https://doi.org/10.1016/j.jinsphys.2005.09.008
- Laing, J. E. & Corrigan, J. E. (1995) Diapause induction and post-diapause emergence in *Trichogramma minutum* Riley (Hymenoptera: Trichogrammatidae): the role of host species, temperature, and photoperiod. *The Canadian Entomologist* 127(1), 103-110. https://doi.org/10.4039/Ent127103-1
- Lessard, E. & Boivin, G. (2013) Effect of low temperature on emergence, fecundity, longevity and host-feeding by *Trichogramma brassicae. BioControl* 58, 319-329. https://doi.org/10.1007/s10526-012-9493-8
- Lopez Jr, J. D. & Morrison, R. K. (1980) Overwintering of *Trichogramma pretiosum* in central Texas. *Environmental Entomology* 9(1), 75-78.
- Mamay, M. & Dağ, E. (2016) Mass Trapping (Kitlesel Yakalama) Tekniğinin Nar Bahçelerinde Harnup güvesi [Apomyelois (= Ectomyelois) ceratoniae Zell. (Lepidoptera: Pyralidae)] Mücadelesindeki Etkinliği. In II. International Multidisciplinary Congress of Eurasia (Vol. 2, pp. 36-41).
- Mamay, M. (2021) The influence of calyx removal and fruit bagging on carob moth, *Ectomyelois ceratoniae* Zeller (Lepidoptera: Pyralidae), infestation in pomegranate. *Crop Protection*147, 105708. https://doi.org/10.1016/j.cropro.2021.105708
- Mamay, M., Ünlü, L., Yanık, E., Doğramacı, M. & İkinci, A. (2016) Efficacy of mating disruption technique against carob moth, *Apomyelois ceratoniae* Zeller (Lepidoptera: Pyralidae) in pomegranate orchards in Southeast Turkey (Şanlıurfa). *International Journal of Pest Management* 62(4), 295-299. https://doi.org/10.1080/09670874.2016.1185552

- Mnif, I., Elleuch, M., Chaabouni, S. E. & Ghribi, D. (2013) Bacillus subtilis SPB1 biosurfactant: production optimization and insecticidal activity against the carob moth Ectomyelois ceratoniae. Crop Protection 50, 66-72. https://doi.org/10.1016/j.cropro.2013.03.005
- Moezipour, M. (2006) Effect of different temperature and humidity treatments on some biological parameters of two *Trichogramma brassicae* Bezd. populations, collected from pomegranate orchards of Yazd and Saveh. 161 pp. M.Sc. Thesis, Isfahan University of Technology, Iran.
- Moezipour, M., Kafil, M. & Allahyari, H. (2008) Functional response of *Trichogramma brassicae* at different temperatures and relative humidities. *Bulletin of Insectology* 61 (2), 245-250. http://www.bulletinofinsectology.org/Contents/Contentsbullinsect.htm
- Mohamed, H. O. & El-Heneidy, A. H. (2020) Effect of cold storage temperature on quality of the parasitoid, *Trichogrammatoidea bactrae* Nagaraja (Hymenoptera: Trichogrammatidae). *Egyptian Journal of Biological Pest Control* 30(1), 1-13. http://dx.doi.org/10.1186/s41938-020-00288-z
- Muli, B. K., Schulthess, F. & Van den Berg, J. (2010) Performance of Trichogrammatoidea sp. nr lutea Girault (Hymenoptera: Trichogrammatidae) collected from *Mussidia* spp. in Kenya on eggs of six lepidopteran hosts. *Journal of Applied Entomology* 134(6), 521-530. https://doi.org/10.1111/j.1439-0418.2009.01474.x
- Nazeri, M., Ashouri, A. & Hosseini, M. (2015) Can Wolbachia infection improve qualitative characteristics of Trichogramma brassicae reared on cold stored eggs of the host?. International Journal of Pest Management 61(3), 243-249. https://doi.org/10.1080/09670874.2015.1042943
- Olkowski, W. & Zhang, A. (1990) *Trichogramma*: a modern day frontier in biological control. *The IPM practitioner* (USA).
- Otto, M. & Mackauer, M. (1998) The developmental strategy of an idiobiont ectoparasitoid, *Dendrocerus carpenteri*: influence of variations in host quality on offspring growth and fitness. *Oecologia*, 117, 353-364. https://doi.org/10.1007/s004420050668
- Ozder, N. & Sağlam, Ö. (2005) Overwintering of the egg parasitoids *Trichogramma brassicae* and *T. cacoeciae* (Hymenoptera: Trichogrammatidae) in the Thrace region of Turkey. *Journal of Pest Science* 78, 129-132. https://doi.org/10.1007/s10340-005-0083-8
- Ozder, N. (2004) Effect of different cold storage periods on parasitization performance of *Trichogramma cacoeciae* (Hymenoptera, Trichogrammatidae) on eggs of *Ephestia kuehniella* (Lepidoptera, Pyralidae). *Biocontrol Science and Technology* 14(5), 441-447. https://doi.org/10.1080/09583150410001683529
- Pintureau, B. & Daumal, J. (1995) Effects of diapause and host species on some morphometric characters in *Trichogramma* (Hym.: Trichogrammatidae). *Experientia*, 51(1), 67-72. https://doi.org/10.1007/BF01964922
- Pitcher, S. A., Hoffmann, M. P., Gardner, J., Wright, M. G. & Kuhar, T. P. (2002) Cold storage of *Trichogramma ostriniae* reared on *Sitotroga cerealella* eggs. *BioControl* 47, 525-535. https://doi.org/10.1023/A:1016533116798
- **R Core Team** (2020) *R: a language and environment for statistical computing.* R Foundation for Statistical Computing, Vienna.
- Rahimi-Kaldeh, S., Ashouri, A. & Bandani, A. (2017) Long-term storage of sexual and asexual *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae). *Biocontrol Science and Technology* 27(11), 1339-1347. https://doi.org/10.1080/09583157.2017.1397599
- Rivers, D. B., Lee Jr, R. E. & Denlinger, D. L. (2000) Cold hardiness of the fly pupal parasitoid *Nasonia vitripennis* is enhanced by its host *Sarcophaga crassipalpis*. *Journal of Insect Physiology* 46(1), 99-106. https://doi.org/10.1016/S0022-1910(99)00106-7
- Rundle, B. J., Thomson, L. J. & Hoffmann, A. A. (2004) Effects of cold storage on field and laboratory performance of *Trichogramma carverae* (Hymenoptera: Trichogrammatidae) and the response of three *Trichogramma* spp.(*T. carverae*, *T.* nr. *brassicae*, and *T. funiculatum*) to cold. *Journal of Economic Entomology* 97 (2), 213-221. https://doi.org/10.1093/jee/97.2.213
- Tezze, A. A. & Botto, E. N. (2004) Effect of cold storage on the quality of *Trichogramma nerudai* (Hymenoptera: Trichogrammatidae). *Biological control* 30(1), 11-16. https://doi.org/10.1016/j.biocontrol.2003.09.008
- Vaez, N., Pourgholi, Z. & Mohammadi, D. (2018) Influences of cold storage period of Anagasta kuehnieela (Zeller) eggs on biological parameters of Trichogramma brassicae Bezdenko. Journal of Applied Research in Plant Protection 7 (2), 145-162.

- Van den Assem, J. (1971) Some experiments on sex ratio and sex regulation in the pteromalid *Lariophagus distinguendus*. *Netherlands Journal of Zoology* 21, 373-402. https://doi.org/10.1163/002829671X00050
- Van Lenteren, J. C. V. & Tommasini, M. (2002) Mass production, storage, shipment and quality control of natural enemies. In: Albajes R., Gullino, M.L., van Lenteren J. C., & Elad, Y. (eds.) Integrated pest and disease management in greenhouse crops. Springer, Amsterdam, pp 276-294. http://doi.org/10.1007/0-306-47585-5_20
- Wang, Z. & Smith, S. M. (1996) Phenotypic differences between thelytokous and arrhenotokous *Trichogramma minutum* from Zeiraphera canadensis. *Entomologia Experimentalis et Applicata* 78 (3), 315-323. https://doi.org/10.1111/j.1570-7458.1996.tb00796.x
- Warner, R. L., Barnes, M. M. & Laird, E. F. (1990) Reduction of insect infestation and fungal infection by cultural practice in date gardens. *Environmental Entomology* 19 (5), 1618-1623.
- Ziaoddini, Z., Goldansaz, h., Ashouri, A. & Ghasempour, A. (2011) Study on the sexual behavior and male cross-attraction among three geographical populations of the carob moth, *Ectomyelois ceratoniae* under laboratory conditions. *Iranian Journal of Plant Protection Science* 42 (1), 151-161. https://dorl.net/dor/20.1001.1.20084781.1390.42.1.15.1
- Zougari, S., Attia, S., Zouba, A. & Lebdi-Grissa, K. (2021) Effectiveness of mass trapping and *Trichogramma cacoeciae* (Hymenoptera: Trichogrammatidae) releases against *Ectomyelois ceratoniae* (Lepidoptera: Pyralidae) in Tunisian oases. *Biologia* 76, 1175-1188. https://doi.org/10.2478/s11756-020-00628-3

Trichogramma brassicae (Hymenoptera: Trichogrammatidae) تاثير ذفيرهسازى در سرما بر عملكرد Ectomyelois ceratoniae و Ephestia kuehniella

عالیه امینی (💩 مجتبی حسینی (២ ، حسین صادقی (២ و سید حسین گلدانساز ۲ ២

۱- گروه گیاهپزشکی، دانشکده کشاورزی، دانشگاه فردوسی مشهد، مشهد، ایران *amini.alich@yahoo.com* ^{(b} h*ttps://orcid.org/0009-0006-8142-9475* ﷺ) m.hosseini@um.ac.ir ^b h*ttps://orcid.org/0000-0001-6708-4657* ﷺ sadeghin@um.ac.ir ^c h*ttps://orcid.org/0000-0002-8329-2699* ۲- گروه گیاهپزشکی، دانشکده علوم و مهندسی کشاورزی، پردیس کشاورزی و منابع طبیعی دانشگاه تهران، کرج، ایران goldansz@ut.ac.ir ^c h*ttps://orcid.org/0000-0001-7574-7553*

تاريخچه مقاله

دريافت: ١٤٠٢/٠٧/٠٩ | پذيرش: ١٤٠٢/١١/٠١ | دبير تخصصي: يعقوب فتحي پور

مٍكيده

زنبورهای پارازیتویید خانواده Trichogrammatidae مهمترین دشمنان طبیعی در برنامه های کنترل بیولوژیک تعداد زیادی از آفات مهم بالپولکدار در سرتاسر دنیا هستند. از این رو وجود روش های مناسب نگهداری برای این پارازیتوییدها در کنترل بیولوژیک ارزشمند است. در این تحقیق عملکرد زنبور *Trichogramma brassic پرو*ش یافته روی دو میزبان مختلف و دو دوره متفاوت ذخیره سازی در سرما بررسی شد. شفیره زنبورهایی که روی تخم بید آرد *Ephestia kuchniella* و شب پره کرم گلوگاه انار *Ectomyelols* روی نوم میزبان مختلف و دو دوره متفاوت ذخیره سازی در سرما بررسی شد. شفیره زنبورهایی که روی تخم بید آرد *Aubernice ای و می*زبان مختلف و دو دوره متفاوت ذخیره سازی در سرما براسی شد. شفیره زنبورهای از ۲۰۱۹ می می می از ترجم می از ترکیب تیماری روی نوم زنبورهای نورش یافته بودند، به مدت ۳۰ روز در دمای ۱۰ درجه سلسیوس و سپس به مدت ۴ و ۸ هفته در دمای ۴ درجه سلسیوس نگهداری شدند. اثر ترکیب تیماری روی نرخ ظهور زنبورها، نسبت جنسی و تعداد زنبورهای ماده و نر در نسل مادری (G) و نتاج آنها (F۱) بررسی شد. به علاوه، میزان بد شکلی زنبورها و نرخ پارازیتیسم به ترتیب در سال مادری و نتاج تحت تاثیر ترکیب تیماری اندازه گیری شد. نتایج این مطالعه نشان داد که نرخ ظهور زنبور *Thichogramic در* (F0) به طرو می میزان و مدت ذکیره سازی در سرما بود. راماده به نر) زنبورهای و و دارج ای ای دادره تری مداناه نتاین مطالعه نشان داد که نرخ ظهور زنبور و تیم ۸ هفته در در میزبان و مدت ذخیره سازی در سرما بود. حداکثر و حداقل نرخ ظهور به ترتیب در *Cetatoniae و تیم* ۸ هفته ذخیره سازی در سرما بود، اما بر همکنش عوامل مذکور روی این ویژگی زنبورها در می از نرد و می بان با افزایش مدت زمان (ماده به نر) زنبورهای و 10 می بود. حداکثر و حداقل نرخ ظهور به ترتیب در معاد و تیمار ۸ هفته ذخیره سازی در سرما بود. معنیدار نشد. همچنین بیبالی و بدشکلی در و ۲۰ می باین یا طول دوره ذخیره سازی در سرما بود، اما بر همکنش عوامل مذکور روی این ویژگی زنبورها مدن بر می باین با افزایش مدت زمان راماده به نری زنبورهای و 1 به مورت معنی داری تحت تاثیر هیچیک از تیمارها قرار نگرفت. نرخ پارازیتسیم زنبورهای برورش یافته روی هر دوری تامه رای مدر می به در بر باره در سرما بود، اما بر همکنش عوامل مذکور روی این ویژگی زنبورها دوی قدرمان معنیدار نشد. همچن

كلمات كليدى: كنترل بيولژيك، پارازيتوييد تخم، رهاسازى اشباعى، كنترل كيفيت، شب پره كرم گلوگاه انار

نویسنده مسئول: مجتبی حسینی (پست الکترونیک: m.hosseini@um.ac.ir)

Citation: Amini, A., Hosseini, M., Sadeghi, H. & Goldansaz, S. H. (2024) Effect of cold storage on performance of *Trichogramma brassicae* (Hymenoptera: Trichogrammatidae) reared on *Ephestia kuehniella* and *Ectomyelois ceratoniae*. *J. Entomol. Soc. Iran* 44 (2), 129-138. https://doi.org/10.61186/jesi.44.2.2